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A community perspective on the concept of marine holobionts: current status, challenges, and future directions

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page 19

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ABSTRACT

Host-microbe interactions play crucial roles in marine ecosystems. However, we still have very little understanding of the mechanisms that govern these relationships, the evolutionary processes that shape them, and their ecological consequences. The holobiont concept is a renewed paradigm in biology that can help to describe and understand these complex systems. It posits that a host and its associated microbiota with which it interacts, form a holobiont, and have to be studied together as a coherent biological and functional unit to understand its biology, ecology, and evolution. Here we discuss critical concepts and opportunities in marine holobiont research and identify key challenges in the field. We highlight the potential economic, sociological, and environmental impacts of the holobiont concept in marine biological, evolutionary, and environmental sciences. Given the connectivity and the unexplored biodiversity specific to marine ecosystems, a deeper understanding of such complex systems requires further technological and conceptual advances, e.g., the development of controlled experimental model systems for holobionts from all major lineages and the modeling of (info)chemical-mediated interactions between organisms. Here we propose that one significant challenge is to bridge cross-disciplinary research on tractable model systems in order to address key ecological and evolutionary questions. This first step is crucial to decipher the main drivers of the dynamics and evolution of holobionts and to account for the holobiont concept in applied areas, such as the conservation, management, and exploitation of marine ecosystems and resources, where practical solutions to predict and mitigate the impact of human activities are more important than ever.

Subjects Ecology, Marine Biology, Microbiology

Keywords Evolution, Ecosystem services, Symbiosis, Host-microbiota interactions, Marine holobionts, Dysbiosis

RATIONALE, INTENDED AUDIENCE, AND SURVEY METHODOLOGY

The idea of considering organisms in connection with the complex microbial communities they are associated with is a concept rapidly gaining in importance in a wide field of life and environmental sciences. It goes along with an increasing awareness that many organisms depend on complex interactions with their symbiotic microbiota for different aspects of their life, even though the extent of dependencies may vary strongly ([Hammer, Sanders & Fierer, 2019](#)). The host and its associated microbiota are considered a single ecological unit, the holobiont. This implies a real paradigm shift. Marine environments harbor most of the diversity of life in terms of the number of lineages that coexist, and the constant presence of surrounding water as a potential carrier of metabolites but also microbes facilitates tight interactions between these lineages, making a “holistic” view of these environments and the organisms that inhabit them particularly important.

This paper is intended for both scientists seeking an overview of recent developments in marine holobiont research and as a reference for policymakers. We review the state of the art in the field research and identify key challenges, possible solutions, and opportunities in the field. Our work is based on the result of a foresight workshop hosted in March 2018, which brought together an interdisciplinary group of 31 scientists. These scientists were selected for their complementary expertise in philosophy, evolution, computer sciences, marine biology, ecology, chemistry, microscopy, and microbiology, as well as for their work with a wide range of different model systems from phytoplankton, via macroalgae, corals, and sponges, to bacterial communities of hydrothermal vents. After a three-day brainstorming session, the resulting ideas and discussions were divided into different topics, and groups of two or more scientists were appointed to draft each section, each based on their expertise in the field, their knowledge of the literature, and literature searches. The assembled paper was then corrected and completed by the entire consortium.

MARINE HOLOBIONTS FROM THEIR ORIGINS TO THE PRESENT

The history of the holobiont concept

Holism is a philosophical notion first proposed by [Aristotle](#) in the 4th century BC. It states that systems should be studied in their entirety, with a focus on the interconnections between their various components rather than on the individual parts (Met. Z.17, 1041b11–33). Such systems have **emergent properties** that result from the behavior of a system that is “larger than the sum of its parts”. However, a major shift away from holism occurred during the Age of “Enlightenment” when the dominant thought summarized as “dissection science” was to focus on the smallest component of a system as a means of understanding it.

The idea of holism started to regain popularity in biology when the endosymbiosis theory was first proposed by [Mereschkowski \(1905\)](#) and further developed by [Wallin \(1925\)](#). Still accepted today, this theory posits a single origin for eukaryotic cells through the **symbiotic** assimilation of prokaryotes to form first mitochondria and later plastids (the latter through several independent symbiotic events) via **phagocytosis** (reviewed in [Archibald, 2015](#)). These ancestral and founding symbiotic events, which prompted the metabolic and cellular complexity of eukaryotic life, most likely occurred in the ocean ([Martin et al., 2008](#)).

Despite the general acceptance of the endosymbiosis theory, the term “holobiosis” or “**holobiont**” did not immediately enter the scientific vernacular. It was coined independently by [Meyer-Abich \(1943\)](#) ([Baedke, Fábregas-Tejeda & Nieves Delgado, 2020](#)) and by [Lynn Margulis](#) in 1990, who proposed that evolution has worked mainly through symbiosis-driven leaps that merged organisms into new forms, referred to as “holobionts”, and only secondarily through gradual mutational changes ([Margulis & Fester, 1991](#); [O’Malley, 2017](#)). However, the concept was not widely used until it was co-opted by coral biologists over a decade later. Corals and dinoflagellate algae of the family Symbiodiniaceae are one of the most iconic examples of symbioses found in nature; most corals are incapable of long-term survival without the products of photosynthesis provided by their

endosymbiotic algae. [Rohwer et al. \(2002\)](#) were the first to use the word “holobiont” to describe a unit of selection *sensu* Margulis ([Rosenberg et al., 2007b](#)) for corals, where the holobiont comprised the cnidarian polyp (**host**), algae of the family Symbiodiniaceae, various ectosymbionts (endolithic algae, prokaryotes, fungi, other unicellular eukaryotes), and viruses.

Although initially driven by studies of marine organisms, much of the research on the emerging properties and significance of holobionts has since been carried out in other fields of research: the **microbiota** of the rhizosphere of plants or the animal gut became predominant models and have led to an ongoing paradigm shift in agronomy and medical sciences ([Bulgarelli et al., 2013](#); [Shreiner, Kao & Young, 2015](#); [Faure, Simon & Heulin, 2018](#)). Holobionts occur in terrestrial and aquatic habitats alike, and several analogies between these ecosystems can be made. For example, in all of these habitats, interactions within and across holobionts such as induction of chemical defenses, nutrient acquisition, or biofilm formation are mediated by chemical cues and signals in the environment, dubbed **infochemicals** ([Loh et al., 2002](#); [Harder et al., 2012](#); [Rolland et al., 2016](#); [Saha et al., 2019](#)). Nevertheless, we can identify two major differences between terrestrial and aquatic systems. First, the physicochemical properties of water result in higher chemical connectivity and signaling between macro- and micro-organisms in aquatic or moist environments. In marine ecosystems, carbon fluxes also appear to be swifter and trophic modes more flexible, leading to higher plasticity of functional interactions across holobionts ([Mitra et al., 2013](#)). Moreover, dispersal barriers are usually lower, allowing for faster microbial community shifts in marine holobionts ([Kinlan & Gaines, 2003](#); [Burgess et al., 2016](#); [Martin-Platero et al., 2018](#)). Secondly, phylogenetic diversity at broad taxonomic scales (i.e., supra-kingdom, kingdom and phylum levels), is higher in aquatic realms compared to land, with much of the aquatic diversity yet to be uncovered ([De Vargas et al., 2015](#); [Thompson et al., 2017](#)), especially marine viruses ([Middelboe & Brussaard, 2017](#); [Gregory et al., 2019](#)). The recent discovery of such astonishing marine microbial diversity in parallel with the scarcity of marine holobiont research suggest a high potential for complex cross-lineage interactions yet to be explored in marine holobionts ([Fig. 1](#)).

The boundaries of holobionts are usually delimited by a physical gradient, which corresponds to the area of local influence of the host, e.g., in unicellular algae the so-called **phycosphere** ([Seymour et al., 2017](#)). However, they may also be defined in a context-dependent way as a “Russian Matryoshka doll”, setting the boundaries of the holobiont depending on the interactions and biological functions that are being considered. Thus holobionts may encompass all levels of host-symbiont associations from intimate **endosymbiosis** with a high degree of co-evolution up to the community and ecosystem level; a concept referred to as “**nested ecosystems**” ([Fig. 2](#); [McFall-Ngai et al., 2013](#); [Pita et al., 2018](#)).

Such a conceptual perspective raises fundamental questions not only regarding the interaction between the different components of holobionts and processes governing their dynamics, but also of the relevant units of selection and the role of co-evolution. For instance, plant and animal evolution involves new functions co-constructed by members of the holobiont or elimination of functions redundant among them ([Selosse, Bessis &](#)

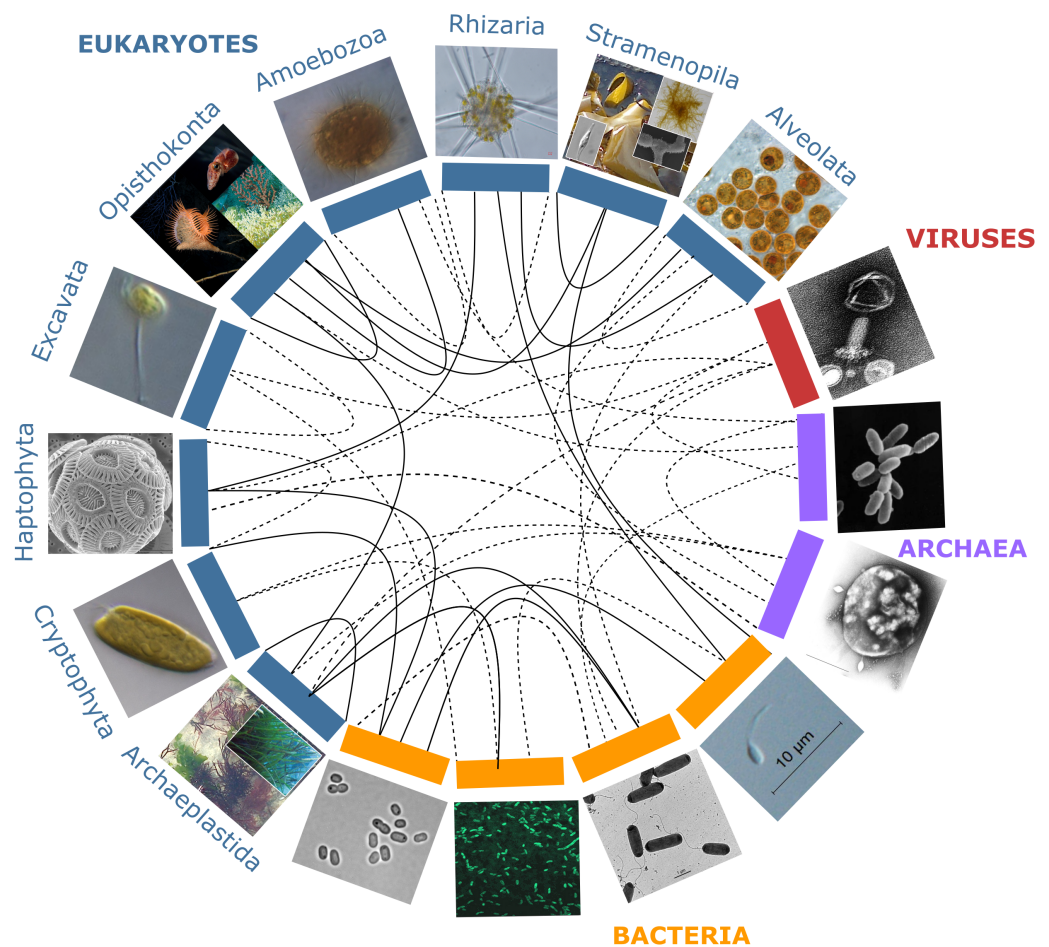


Figure 1 Partners forming marine holobionts. They are widespread across the tree of life including all kingdoms (eukaryotes, bacteria, archaea, viruses), and represent a large diversity of potential models for exploring complex biotic interactions across lineages. Plain lines correspond to holobionts referred to in the present manuscript. Dashed lines are examples of potential interactions. Photo credits: Archaeplastida –Catherine Leblanc, Ulisse Cardini; Excavata - Roscoff Culture Collection (<http://roscoff-culture-collection.org/rcc-strain-details/1065>), Attribution 4.0 International (CC BY 4.0); Amoebozoa - Roscoff Culture Collection (<http://roscoff-culture-collection.org/rcc-strain-details/1067>), Attribution 4.0 International (CC BY 4.0); Cryptophyta –Roscoff Culture Collection (<http://roscoff-culture-collection.org/rcc-strain-details/1998>), Attribution 4.0 International (CC BY 4.0); Stramenopila –Catherine Leblanc, Simon M Dittami; Alveolata –Allison Lewis (<https://commons.wikimedia.org/wiki/File:Symbiodinium.png>), Creative Commons Attribution-Share Alike 4.0 International license; Rhizaria –Fabrice Not; Haptophyta –Alison R. Taylor ([https://en.wikipedia.org/wiki/Emiliania_huxleyi#/media/File:Emiliania_huxleyi_coccolithophore_\(PLOS\).png](https://en.wikipedia.org/wiki/Emiliania_huxleyi#/media/File:Emiliania_huxleyi_coccolithophore_(PLOS).png)), Attribution 2.5 Generic (CC BY 2.5); Opisthokonta –HeikeM (https://fr.wikipedia.org/wiki/R%C3%A9cif_corallien_d%27eau_froide#/media/Fichier:Joon1.jpg, Public Domain), NOAA Photo Library (https://en.wikipedia.org/wiki/Sea_anemone#/media/File:Actinoscyphia_aurelia_1.jpg, Public Domain), Squid (Chris Frazee, Margaret McFall-Ngai, https://en.wikipedia.org/wiki/Squid#/media/File:Euprymna_scolopes_-_image.pbio.v12.i02.g001.png, Attribution 4.0 International (CC BY 4.0)); Bacteria –Marinobacter (Astrid Gärdes, Eva Kaepfel, Aamir Shehzad, Shalin Seebah, Hanno Teeling, Pablo Yarza, Frank Oliver Glöckner, Hans-Peter Grossart, Matthias S. Ullrich, <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3035377/figure/f1/>, Attribution 2.5 Generic (CC BY 2.5)), (continued on next page...)

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Figure 1 (...continued)

Synechococcus (Masur, https://en.wikipedia.org/wiki/Synechococcus#/media/File:Synechococcus_PCC_7002_BF.jpg, Public Domain), *Vibrio fischeri* (Alan Cann, <https://www.flickr.com/photos/ajc1/252308050/>, Attribution-NonCommercial 2.0 Generic (CC BY-NC 2.0)), *Hyphomonas* - Holomarine consortium (Simon M Dittami); *Archaea* - *Halobacterium* (NASA, <https://commons.wikimedia.org/wiki/File:Halobacteria.jpg>, Public Domain), *Sulfolobus* (Xiangyux, <https://de.wikipedia.org/wiki/Archaeen#/media/File:RT8-4.jpg>, Public Domain); Viruses - Matthew B Sullivan, Maureen L Coleman, Peter Weigle, Forest Rohwer, Sallie W Chisholm (<https://en.wikipedia.org/wiki/Cyanophage#/media/File:Cyanophages.png>), Attribution 2.5 Generic (CC BY 2.5).

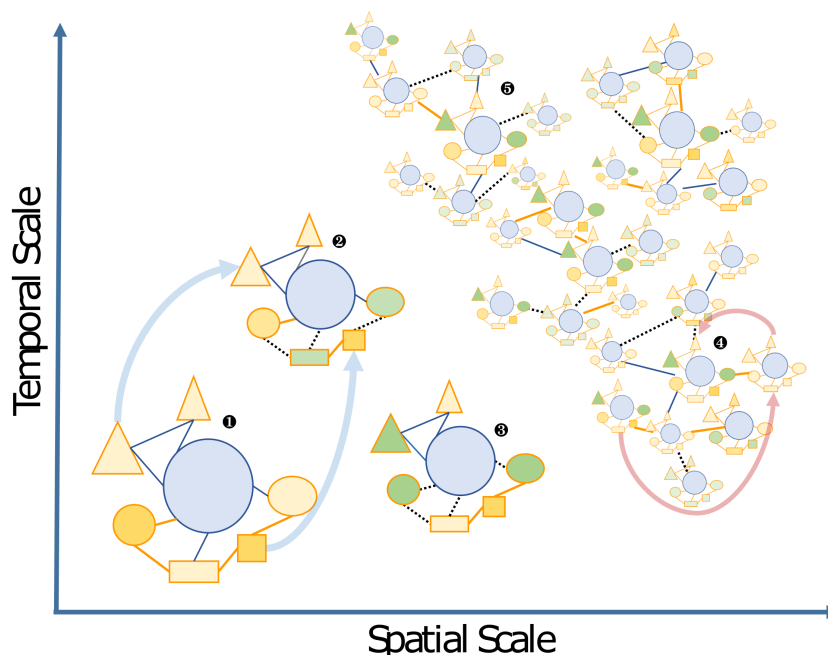


Figure 2 Schematic view of the “Russian Doll” complexity and dynamics of holobionts, according to diverse spatiotemporal scales. The host (blue circles), and associated microbes (all other shapes) including bacteria and eukaryotes that may be inside (i.e., endosymbiotic) or outside the host (i.e., ectosymbiotic) are connected by either beneficial (solid orange lines), neutral (solid blue lines) or pathogenic (dashed black lines) interactions, respectively. Changes from beneficial or neutral to pathogenic interactions are typical cases of dysbiosis. The different clusters can be illustrated by the following examples: 1, a model holobiont in a stable physiological condition (e.g., in controlled laboratory condition); 2 and 3, holobionts changing during their life cycle or subjected to stress conditions—examples of vertically transmitted microbes are indicated by light blue arrows; 4 and 5, marine holobionts in the context of global sampling campaigns or long-term time series—examples of horizontal transmission of microbes and holobionts are illustrated by pink arrows.

Full-size [DOI: 10.7717/peerj.10911/fig-2](https://doi.org/10.7717/peerj.10911/fig-2)

Pozo, 2014), and it is likely that these processes are also relevant in marine holobionts. *Rosenberg et al. (2010)* and *Rosenberg & Zilber-Rosenberg (2018)* argued that all animals and plants can be considered holobionts, and thus advocate the **hologenome** theory of evolution, suggesting that natural selection acts at the level of the holobiont and its hologenome. This interpretation of Margulis’ definition of a “holobiont” considerably broadened fundamental concepts in evolution and speciation and has not been free of

criticism (Douglas & Werren, 2016), especially when applied at the community or ecosystem level (Moran & Sloan, 2015). More recently, it has been shown that species that interact indirectly with the host can also be important in shaping coevolution within mutualistic multi-partner assemblages (Guimarães et al., 2017). Thus, the holobiont concept and the underlying complexity of holobiont systems should be better defined and further considered when addressing evolutionary and ecological questions.

Marine holobiont models

Today, an increasing number of marine model organisms, both unicellular and multicellular, are being used in holobiont research (Fig. 1), often with different emphasis and levels of experimental control, but altogether covering a large range of scientific topics. Here, we provide several illustrative examples of this diversity and some of the insights they have provided, distinguishing between “environmental models”, chosen for their environmental, evolutionary, economical, or ecological importance, or for historical reasons, but in which microbiome composition is not or only partially controlled, and “controlled bi- or trilateral associations”, which can be kept separately from their symbionts under laboratory conditions and are particularly useful to develop functional approaches and study the mechanisms of symbiotic interactions.

Environmental models: Within the animal kingdom, and in addition to corals and sponges, which will be discussed below, the discovery of deep-sea hydrothermal vents revealed symbioses of animals with chemosynthetic bacteria that have later been found in many other marine ecosystems (Dubilier, Bergin & Lott, 2008; Rubin-Blum et al., 2019) and frequently exhibit high levels of metabolic and taxonomic diversity (Duperron et al., 2008; Petersen et al., 2016; Ponnudurai et al., 2017). In the SAR supergroup, in addition to well-known models such as diatoms, radiolarians and foraminiferans, both heterotrophic protist dwellers harboring endosymbiotic microalgae, are emerging as ecological models for unicellular photosymbiosis due to their ubiquitous presence in the world’s oceans (Decelle, Colin & Foster, 2015; Not et al., 2016). Among the haptophytes, the cosmopolitan *Emiliania huxleyi*, promoted by associated bacteria (Seyedsayamdost et al., 2011; Segev et al., 2016), produces key intermediates in the carbon and sulfur biogeochemical cycles, making it an important model phytoplankton species. Finally, within the Archaeplastida, the siphonous green alga *Bryopsis* is an example of a model that harbors heterotrophic endosymbiotic bacteria, some of which exhibit patterns of co-evolution with their hosts (Hollants et al., 2013).

Controlled bi- or trilateral associations: Only a few models, covering a small part of the overall marine biodiversity, are currently being cultivated ex-situ and can be used in fully controlled experiments, where they can be cultured **aprosymbiotically**. The flatworm *Symsagittifera* (= *Convoluta*) *roscoffensis* (Arboleda et al., 2018), the sea anemone *Exaiptasia* (Baumgarten et al., 2015; Wolfowicz et al., 2016), the upside-down jellyfish *Cassiopea* (Ohdera et al., 2018), and their respective intracellular green and dinoflagellate algae have, in addition to corals, become models for fundamental research on evolution of metazoan-algal photosymbiosis. In particular, the sea anemone *Exaiptasia* has been used to explore photobiology disruption and restoration of cnidarian symbioses (Lehnert, Burriesci

& Pringle, 2012). The *Vibrio*-squid model provides insights into the effect of microbiota on animal development, circadian rhythms, and immune systems (McFall-Ngai, 2014). The unicellular green alga *Ostreococcus*, an important marine primary producer, has been shown to exchange vitamins with specific associated bacteria (Cooper et al., 2019). The green macroalga *Ulva mutabilis* has enabled the exploration of bacteria-mediated growth and morphogenesis including the identification of original chemical interactions in the holobiont (Wichard, 2015; Kessler et al., 2018). Although the culture conditions in these highly-controlled model systems differ from the natural environment, these systems are essential to gain elementary mechanistic understanding of the functioning, the roles, and the evolution of marine holobionts.

The influence of marine holobionts on ecological processes

Work on model systems has demonstrated that motile and macroscopic marine holobionts can act as dissemination vectors for geographically restricted microbial taxa. Pelagic mollusks or vertebrates are textbook examples of high dispersal capacity organisms (e.g., against currents and through stratified water layers). It has been estimated that fish and marine mammals may enhance the original dispersion rate of their microbiota by a factor of 200 to 200,000 (Troussellier et al., 2017) and marine birds may even act as bio-vectors across ecosystem boundaries (Bouchard Marmen et al., 2017). This host-driven dispersal of microbes can include non-native or invasive species as well as pathogens (Troussellier et al., 2017).

A related ecological function of holobionts is their potential to sustain rare species. Hosts provide an environment that favors the growth of specific microbial communities distinct from the surrounding environment (including rare microbes). They may, for instance, provide a nutrient-rich niche in the otherwise nutrient-poor surroundings (Smriga, Sandin & Azam, 2010; Webster et al., 2010; Burke et al., 2011a; Burke et al., 2011b; Chiarello et al., 2018).

Lastly, biological processes regulated by microbes are important drivers of global biogeochemical cycles (Falkowski, Fenchel & Delong, 2008; Madsen, 2011; Anantharaman et al., 2016). In the open ocean, it is estimated that symbioses with the cyanobacterium UCYN-A contribute ~20% to total N₂ fixation (Thompson et al., 2012; Martínez-Pérez et al., 2016). In benthic systems, sponges and corals may support entire ecosystems via their involvement in nutrient cycling thanks to their microbial partners (Raina et al., 2009; Fiore et al., 2010; Cardini et al., 2015; Pita et al., 2018), functioning as sinks and sources of nutrients. In particular the “sponge loop” recycles dissolved organic matter and makes it available to higher trophic levels in the form of detritus (De Goeij et al., 2013; Fiore et al., 2010; Rix et al., 2017). In coastal sediments, bivalves hosting methanogenic archaea have been shown to increase the benthic methane efflux by a factor of up to eight, potentially accounting for 9.5% of total methane emissions from the Baltic Sea (Bonaglia et al., 2017). Such impressive metabolic versatility is accomplished because of the simultaneous occurrence of disparate biochemical machineries (e.g., aerobic and anaerobic pathways) in individual symbionts, providing new metabolic abilities to the holobiont, such as the synthesis of specific essential amino acids, photosynthesis, or chemosynthesis (Dubilier,

Bergin & Lott, 2008; Venn, Loram & Douglas, 2008). Furthermore, the interaction between host and microbiota can potentially extend the metabolic capabilities of a holobiont in a way that augments its resilience to environmental changes (*Berkelmans & Van Oppen, 2006; Gilbert et al., 2010; Dittami et al., 2016; Shapira, 2016; Godoy et al., 2018*), or allow it to cross biotope boundaries (e.g., *Woyke et al., 2006*) and colonize extreme environments (*Bang et al., 2018*). Holobionts thus contribute to marine microbial diversity and possibly resilience in the context of global environmental changes (*Troussellier et al., 2017*) and it is paramount to include the holobiont concept in predictive models that investigate the consequences of human impacts on the marine realm and its biogeochemical cycles.

CHALLENGES AND OPPORTUNITIES IN MARINE HOLOBIONT RESEARCH

Marine holobiont assembly and regulation

Two critical challenges partially addressed by using model systems are (1) to decipher the factors determining holobiont composition and (2) to elucidate the impacts and roles of the different partners in these complex systems over time. Some marine organisms such as bivalves transmit part of the microbiota maternally (*Bright & Bulgheresi, 2010; Funkhouser & Bordenstein, 2013*). In other marine holobionts, vertical transmission may be weak and inconsistent, whereas mixed **modes of transmission (vertical and horizontal)** or intermediate modes (pseudo-vertical, where horizontal acquisition frequently involves symbionts of parental origin) are more common (*Björk et al., 2019*). Identifying the factors shaping holobiont composition and understanding their evolution is highly relevant for marine organisms given that most marine hosts display a high specificity for their microbiota and even patterns of **phylosymbiosis** (*Brooks et al., 2016; Kazamia et al., 2016; Pollock et al., 2018*), despite a highly connected and microbe-rich environment.

During microbiota transmission (whether vertical or horizontal), *selection* by the host and/or by other components of the microbiome, is a key process in establishing or maintaining a holobiont microbial community that is distinct from the environment. The immune system of the host, e.g., via the secretion of specific antimicrobial peptides (*Franzenburg et al., 2013; Zheng, Liwinski & Elinav, 2020*), is one way of performing this selection in both marine and terrestrial holobionts.

Another way of selecting a holobiont microbial community is by chemically mediated **microbial gardening**. This concept has been demonstrated for land plants, where root exudates manipulate **microbiome** composition (*Lebeis et al., 2015*). In marine environments, the phylogenetic diversity of hosts and symbionts suggests both conserved and marine-specific chemical interactions, but studies are still in their infancy. For instance, seaweeds can chemically garden beneficial microbes, facilitating normal morphogenesis and increasing disease resistance (*Kessler et al., 2018; Saha & Weinberger, 2019*), and seaweeds and corals structure their surface-associated microbiome by producing chemo-attractants and anti-bacterial compounds (*Harder et al., 2012; Ochsenkühn et al., 2018*). There are fewer examples of chemical gardening in unicellular hosts, but it seems highly likely that similar processes are in place (*Gribben et al., 2017; Cirri & Pohnert, 2019*).

In addition to selection, ecological drift, dispersal and evolutionary diversification have been proposed as key processes in community assembly, but are difficult to estimate in microbial communities (Nemergut et al., 2013). The only data currently at our disposal to quantify these processes are the diversity and distribution of microbes. Considering the high connectivity of aquatic environments, differences in marine microbial communities are frequently attributed to a combination of selection and drift, rather than limited dispersal (e.g., Burke et al., 2011a), a conclusion which, in the future, could be refined by conceptual models developed for instance for soil microbial communities (Stegen et al., 2013; Dini-Andreote et al., 2015). Diversification is mainly considered in the sense of coevolution or adaptation to host selection, which may also be driven by the horizontal acquisition of genes. However, cospeciation is challenging to prove (De Vienne et al., 2013; Moran & Sloan, 2015) and only few studies have examined this process in marine holobionts to date, each focused on a restricted number of actors (e.g., Peek et al., 1998; Lanterbecq, Rouse & Eeckhaut, 2010).

Perturbations in the transmission or the recruitment of the microbiota can lead to **dysbiosis**, and eventually microbial infections (Selosse, Bessis & Pozo, 2014; De Lorgeril et al., 2018). Dysbiotic microbial communities are frequently determined by stochastic processes and thus display higher variability in their composition than those of healthy individuals. This observation is in line with the “**Anna Karenina principle**” (Zaneveld, McMinds & Vega Thurber, 2017), although there are exceptions to this rule (e.g., Marzinelli et al., 2015). A specific case of dysbiosis is the so-called “**Rasputin effect**” where benign endosymbionts opportunistically become detrimental to the host due to processes such as reduction in immune response under food deprivation, coinfections, or environmental pressure (Overstreet & Lotz, 2016). Many diseases are now interpreted as the result of a microbial imbalance and the rise of opportunistic or polymicrobial infections upon host stress (Egan & Gardiner, 2016). For instance in reef-building corals, warming destabilizes cnidarian-dinoflagellate associations, and some beneficial *Symbiodiniaceae* strains switch their physiology and sequester more resources for their own growth at the expense of the coral host, leading to coral bleaching and even death (Baker et al., 2018).

Increasing our knowledge on the contribution of these processes to holobiont community assembly in marine systems is a key challenge, which is of particular urgency today in the context of ongoing global change. Moreover, understanding how the community and functional structure of resident microbes are resilient to perturbations remains critical to predict and promote the health of their host and the ecosystem. Yet, the contribution of the microbiome is still missing in most quantitative models predicting the distribution of marine macro-organisms, or additional information on biological interactions would be required to make the former more accurate (Bell et al., 2018).

Integrating marine model systems with large-scale studies

By compiling a survey of the most important trends and challenges in the field of marine holobiont research (Fig. 3), we identified two distinct opinion clusters: one focused on mechanistic understanding and work with model systems whereas another targets large-scale and heterogeneous data set analyses and predictive modeling. This illustrates that, on

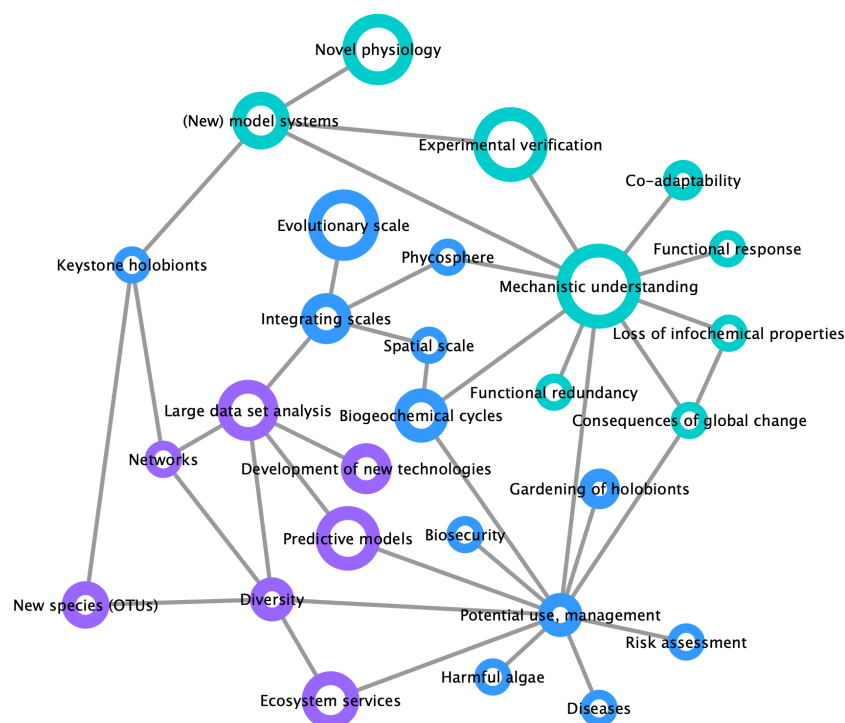


Figure 3 Mind map of key concepts, techniques, and challenges related to marine holobionts. The basis of this map was generated during the Holomarine workshop held in Roscoff in 2018 (<https://www.euromarinetwork.eu/activities/HoloMarine>). The size of the nodes reflects the number of votes each keyword received from the participants of the workshop (total of 120 votes from 30 participants). The two main clusters corresponding to predictive modeling and mechanistic modeling, are displayed in purple and turquoise, respectively. Among the intermediate nodes linking these disciplines (blue) “potential use, management” was the most connected.

Full-size DOI: 10.7717/peerj.10911/fig-3

the one hand, the scientific community is interested in the establishment of models for the identification of specific molecular interactions between marine organisms at a given point in space and time, up to the point of synthesizing functional mutualistic communities in vitro (*Kubo et al., 2013*). On the other hand, another part of the community is moving towards global environmental sampling schemes such as the *TARA* Oceans expedition (*Pesant et al., 2015*) or the Ocean Sampling Day (*Kopf et al., 2015*), and towards long-term data series (e.g., *Wiltshire et al., 2010*; *Harris, 2010*). What emerges as both lines of research progress is the understanding that small-scale functional studies in the laboratory are inconsequential unless made applicable to ecologically-relevant systems. At the same time, and despite the recent advances in community modeling (*Ovaskainen et al., 2017*), hypotheses drawn from large scale-studies remain correlative and require experimental validation of the mechanisms driving the observed processes. We illustrate the importance of integrating both approaches in *Fig. 3*, where the node related to potential applications was perceived as a central hub at the interface between mechanistic understanding and predictive modeling.

A successful example merging both functional and large-scale approaches, are the root nodules of legumes, which harbor nitrogen-fixing bacteria. In this system, the functioning, distribution, and to some extent the evolution of these nodules, are now well understood ([Epihov et al., 2017](#)). The integration of this knowledge into agricultural practices has led to substantial yield improvements (e.g., [Kavimandan, 1985](#); [Alam et al., 2015](#)). In the more diffuse and partner-rich system of mycorrhizal symbioses between plant roots and soil fungi, a better understanding of the interactions has also been achieved *via* the investigation of environmental diversity patterns in combination with experimental culture systems with reduced diversity ([Van der Heijden et al., 2015](#)).

We advocate the implementation of comparable efforts in marine sciences through interdisciplinary research combining physiology, biochemistry, ecology, and computational modeling. A key factor will be the identification and development of tractable model systems for keystone holobionts that allow hypotheses generated by large-scale data sets to be tested in controlled experiments. Such approaches will enable the identification of organismal interaction patterns within holobionts and nested ecosystems. In addition to answering fundamental questions, they will help address the ecological, societal, and ethical issues that arise from attempting to actively manipulate holobionts (e.g., in aquaculture, conservation, and invasion) in order to enhance their resilience and protect them from the impacts of global change ([Llewellyn et al., 2014](#)).

Emerging methodologies to approach the complexity of holobiont partnerships

As our conceptual understanding of the different levels of holobiont organization evolves, so does the need for multidisciplinary approaches and the development of tools and technologies to handle the unprecedented amount of data and their integration into dedicated ecological and evolutionary models. Here, progress is often fast-paced and provides exciting opportunities to address some of the challenges in holobiont research.

A giant technological stride has been the explosion of affordable “omics” technologies allowing molecular ecologists to move from metabarcoding (i.e., sequencing of a taxonomic marker) to metagenomics or single-cell genomics, metatranscriptomics, and metaproteomics, thus advancing our research from phylogenetic analyses of the holobiont to analyses capable of making predictions about the functions carried out by different components of the holobiont ([Bowers, Doud & Woyke, 2017](#); [Meng et al., 2018](#); [Fig. 4](#)). These approaches are equally useful in marine and in terrestrial environments, but the scarcity of well-studied lineages in the former makes the generation of good annotations and reference databases challenging for marine biologists. Metaproteomics combined with stable isotope fingerprinting can help study the metabolism of single lineages within the holobiont ([Kleiner et al., 2018](#)). In parallel, meta-metabolomics approaches have advanced over the last decades, and can be used to unravel the chemical interactions between partners. One limitation particularly relevant to marine systems is that many compounds are often not referenced in the mostly terrestrial-based databases, although recent technological advances such as molecular networking and meta-mass shift chemical profiling to identify relatives of known molecules may help to overcome this challenge ([Hartmann et al., 2017](#)).

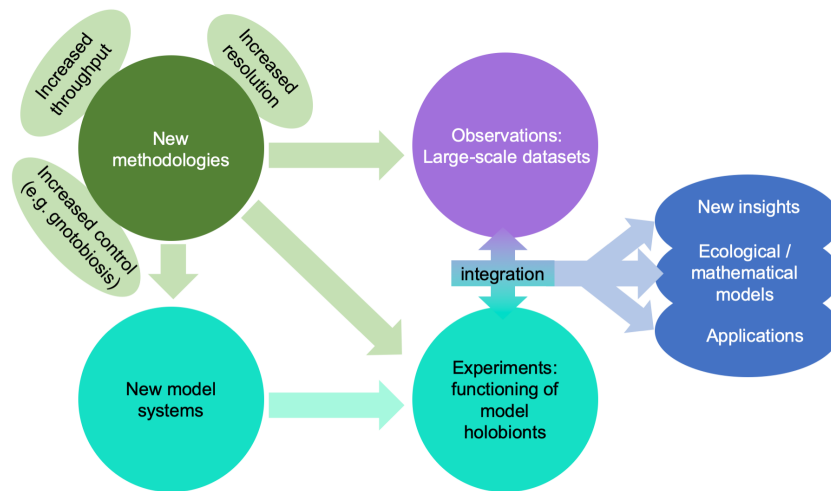


Figure 4 Impact of emerging methodologies (light green) on the main challenges in marine holobiont research identified in this paper (blue). Turquoise and purple correspond to the two main clusters of activity identified in Fig. 3.

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A further challenge in holobiont research is to identify the origin of compounds among the different partners of the holobionts and to determine their involvement in the maintenance and performance of the holobiont system. Well-designed experimental setups may help answer some of these questions (e.g., [Quinn et al., 2016](#)), but they will also require high levels of replication in order to represent the extensive intra-species variability found in marine systems. Recently developed in vivo and in situ imaging techniques combined with metabolomics can provide small-scale spatial and qualitative information (origin, distribution, and concentration of a molecule or nutrient), shedding new light on the contribution of each partner of the holobiont system at the molecular level (e.g., [Geier et al., 2020](#)). The combination of stable isotope labelling and chemical imaging (mass spectrometry imaging such as secondary ion mass spectrometry and matrix-assisted laser desorption ionization, and synchrotron X-ray fluorescence) is particularly valuable in this context, as it enables the investigation of metabolic exchange between the different compartments of a holobiont ([Musat et al., 2016](#); [Raina et al., 2017](#)). Finally, three-dimensional electron microscopy may help evaluate to what extent different components of a holobiont are physically integrated ([Colin et al., 2017](#); [Decelle et al., 2019](#)), where high integration is one indication of highly specific interactions. All of these techniques can be employed in both marine and terrestrial systems, but in marine systems the high phylogenetic diversity of organisms adds to the complexity of adapting and optimizing these techniques.

One consequence of the development of such new methods is the feedback they provide to improve existing models or to develop entirely new ones, e.g., by conceptualizing holobionts as the combination of the interactions between the host and its microbiota ([Skillings, 2016](#); [Berry & Loy, 2018](#)), or by redefining boundaries between the holobiont and its environment ([Zengler & Palsson, 2012](#)). Such models may incorporate metabolic

complementarity between different components of the holobiont ([Dittami, Eveillard & Tonon, 2014](#); [Bordron et al., 2016](#)), e.g., enabling the prediction of testable metabolic properties depending on holobiont composition ([Burgunter-Delamare et al., 2020](#)), or simulate microbial communities starting from different cohorts of randomly generated microbes for comparison with actual metatranscriptomics and/or metagenomics data ([Coles et al., 2017](#)).

A side-effect of these recent developments has been to move holobiont research away from laboratory culture-based experiments. We argue that maintaining or even extending cultivation efforts, possibly via the implementation of “culturomics” approaches as successfully carried out for the human gut microbiome ([Lagier et al., 2012](#)), remains essential to capture the maximum holobiont biodiversity possible and will facilitate the experimental testing of hypotheses and the investigation of physiological mechanisms. A striking example of the importance of laboratory experimentation is the way germ-free mice re-inoculated with cultivated bacteria (the so-called **gnotobiotic** mice) have contributed to the understanding of interactions within the holobiont in animal health, physiology, and behavior (e.g., [Neufeld et al., 2011](#); [Faith et al., 2014](#); [Selosse, Bessis & Pozo, 2014](#)). In terms of gnotobiotic marine organisms there are several examples of microalgae that can be cultured axenically, but gnotobiotic multicellular organisms are rare. One example is the green alga *Ulva mutabilis*, which can be rendered axenic based on the movement of its spores and has been used to study the effects of bacteria-produced morphogens ([Spoerner et al., 2012](#)). There are also examples of gnotobiotic marine fish and mollusks ([Marques et al., 2006](#)). However, in many cases, not all associated microorganisms can be controlled leaving researchers with aposymbiotic cultures (i.e., cultures without the main symbiont(s), as e.g., for the sea anemone *Exaiptasia*) ([Lehnert et al., 2014](#)). Innovations in cultivation techniques for axenic (or germ-free) hosts or in microbial cultivation such as microfluidic systems (e.g., [Pan et al., 2011](#)) and cultivation chips ([Nichols et al., 2010](#)) may provide a way to obtain a wider spectrum of pure cultures. Yet, bringing individual components of holobionts into cultivation can still be a daunting challenge due to the strong interdependencies between organisms as well as the existence of yet unknown metabolic processes that may have specific requirements. In this context, single-cell “omics” analyses can provide critical information on some of the growth requirements of the organisms, and complement approaches of high-throughput culturing ([Gutleben et al., 2018](#)).

Established cultures can then be developed into model systems, e.g., by genome sequencing and the development of genetic tools to move towards mechanistic understanding and experimental testing of hypothetical processes within the holobiont derived from environmental meta “omics” approaches. In this context, CRISPR/cas9 is a particularly promising tool for the genetic modification of both host and symbiont organisms, and has been established for a few marine model systems, including diatoms, cnidarians, annelids, echinoderms, and chordates ([Momose & Concordet, 2016](#)), although this tool has not, to the best of our knowledge, been used so far to decipher host symbiont interactions. “Omics” techniques, coupled to efforts in adapting these genetic tools, have the potential to broaden the range of available models, enabling a better understanding

of the functioning of marine holobionts and their interactions in marine environments (Wichard & Beemelmanns, 2018).

Ecosystem services and holobionts in natural and managed systems

A better understanding of marine holobionts will likely have direct socio-economic consequences for coastal marine ecosystems, estimated to provide services worth almost 50 trillion (10^{12}) US\$ per year (Costanza et al., 2014). Most of the management practices in marine systems have so far been based exclusively on the biology and ecology of macro-organisms. A multidisciplinary approach that provides mechanistic understanding of habitat-forming organisms as holobionts will ultimately improve the predictability and management of coastal ecosystems. For example, host-associated microbiota could be integrated in **biomonitoring** programs as proxies used to assess the health of ecosystems. Microbial shifts and dysbiosis constitute early warning signals that may allow managers to predict potential impacts and intervene more rapidly and effectively (Van Oppen et al., 2017; Marzinelli et al., 2018).

One form of intervention could be to promote positive changes of host-associated microbiota, in ways analogous to the use of pre- and/or probiotics in humans (Singh et al., 2013) or inoculation of beneficial microbes in plant farming (Berruti et al., 2015; Van der Heijden et al., 2015). In macroalgae, beneficial bacteria identified from healthy seaweed holobionts could be used as **biological control** agents and applied to diseased plantlets in order to suppress the growth of bacteria detrimental to the host and to prevent disease outbreaks in aquaculture settings. In addition to bacteria, these macroalgae frequently host endophytic fungi that may have protective functions for the algae (Porrás-Alfaro & Bayman, 2011; Vallet et al., 2018). Host-associated microbiota could also be manipulated to shape key phenotypes in cultured marine organisms. For example, specific bacteria associated with microalgae may enhance algal growth (Amin et al., 2009; Kazamia, Aldridge & Smith, 2012; Le Chevanton et al., 2013), increase lipid content (Cho et al., 2015), and participate in the bioprocessing of algal biomass (Lenneman, Wang & Barney, 2014). More recently, the active modification of the coral microbiota has even been advocated as a means to boost the resilience of the holobiont to climate change (Van Oppen et al., 2015; Peixoto et al., 2017), an approach which would, however, bear a high risk of unanticipated and unintended side effects.

Also, holistic approaches could be implemented in the framework of fish farms. Recent developments including integrated multi-trophic aquaculture, recirculating aquaculture, offshore aquaculture, species selection, and breeding increase yields and reduce the resource constraints and environmental impacts of intensive aquaculture (Klinger & Naylor, 2012). However, the intensification of aquaculture often goes hand in hand with increased amounts of disease outbreaks both in industry and wild stocks. A holistic microbial management approach, e.g., by reducing the use of sterilization procedures and favoring the selection of healthy and stable microbiota consisting of slow-growing K-strategists, may provide an efficient solution to these latter problems, reducing the sensitivity of host to opportunistic pathogens (De Schryver & Vadstein, 2014).

Nevertheless, when considering their biotechnological potential, it should also be noted that marine microbiota are likely vulnerable to anthropogenic influences and that their deliberate engineering, introduction from exotic regions (often hidden in their hosts), or inadvertent perturbations may have profound, and yet entirely unknown, consequences for marine ecosystems. Terrestrial environments provide numerous examples of unwanted plant expansions or ecosystem perturbations linked to microbiota (e.g., [Dickie et al., 2017](#)), and cases where holobionts manipulated by human resulted in pests (e.g., [Clay & Holah, 1999](#)) call for a cautious and ecologically-informed evaluation of holobiont-based technologies in marine systems.

CONCLUSIONS

Marine ecosystems represent highly connected reservoirs of largely unexplored biodiversity. They are of critical importance to feed the ever-growing world population, constitute significant players in global biogeochemical cycles but are also threatened by human activities and global change. In order to unravel some of the basic principles of life and its evolution, and to protect and sustainably exploit marine natural resources, it is paramount to consider the complex biotic interactions that shape the marine communities and their environment. The scope of these interactions ranges from simple molecular signals between two partners, via complex assemblies of eukaryotes, prokaryotes, and viruses with one or several hosts, to entire ecosystems. Accordingly, current key questions in marine holobiont research cover a wide range of topics: What are the exchanges that occur between different partners of the holobiont, and how do they condition their survival, dynamics and evolution? What are the cues and signals driving these exchanges? What are the relevant units of selection and dispersal in marine holobionts? How do holobiont systems and the interactions within them change over time and in different conditions? How do such changes impact ecological processes? How can this knowledge be applied to our benefit and where do we need to draw limits? Identifying and consolidating key model systems while adapting emerging “-omics”, imaging, culturing technologies, and functional analyses via transgenesis (e.g., CRISPR/cas9) to them will be critical to the development of “holobiont-aware” ecosystem models.

The concept of holobionts represents the fundamental understanding that all living organisms have intimate connections with their immediate neighbors, which may impact all aspects of their biology. We believe that this concept of holobionts will be most useful if used with a degree of malleability, enabling us to define units of interacting organisms that are most suitable to answer specific questions. The consideration of the holobiont concept marks a paradigm shift in biological and environmental sciences, but only if scientists work together as an (inter)active and transdisciplinary community bringing together holistic and mechanistic views. This will result in tangible outcomes including a better understanding of evolutionary and adaptive processes, improved modeling of habitats and understanding of biogeochemical cycles, as well as application of the holobiont concept in aquaculture and ecosystem management projects.

Glossary

*	If no other examples of the use of each term are cited below, the definition was based on the online version of the Merriam-Webster dictionary (2019, https://www.merriam-webster.com/) or the Oxford dictionary (2020, https://www.lexico.com/)
Anna Karenina principle	several factors can cause a system to fail, but only a narrow range of parameters characterizes a working system; based on the first sentence of Leo Tolstoy's "Anna Karenina" (1878): "Happy families are all alike; every unhappy family is unhappy in its own way" (Zaneveld, McMinds & Vega Thurber, 2017)
Aposymbiotic culture	a culture of a host or a symbiont without its main symbiotic partner(s) (e.g., Kelty & Cook, 1976). In contrast to gnotobiotic cultures, aposymbiotic cultures are usually not germ-free
Biological control (biocontrol)	methods for controlling diseases or pests by introducing or supporting natural enemies of the former (see e.g., Hoitink & Boehm, 1999)
Biomonitoring	the use of living organisms as quantitative indicator for the health of an environment or ecosystem (Holt & Miller, 2010)
Community assembly process	the processes that shape community composition in a given habitat, according to Nemergut et al. (2013) the four main forces relevant for community assembly are evolutionary diversification, dispersal, selection, and ecological drift
Dysbiosis	microbial imbalance in a symbiotic community that affects the health of the host (Egan & Gardiner, 2016)
Ecological process	the processes responsible for the functioning and dynamics of ecosystems including biogeochemical cycles, community assembly processes, interactions between organisms, and climatic processes (see e.g., Bennett et al., 2009)
Ecosystem services	any direct or indirect benefits that humans can draw from an ecosystem; they include provisioning services (e.g., food), regulating services (e.g., climate), cultural services (e.g., recreation), and supporting services (e.g., habitat formation) (Millennium Ecosystem Assessment Panel, 2005)
Ectosymbiosis	a symbiotic relationship in which symbionts live on the surface of a host. This includes, for instance, algal biofilms or the skin microbiome (Nardon & Charles, 2001)
Emergent property	a property of complex systems (e.g., holobionts), which arises from interactions between the components and that is not the sum of the component properties (see e.g., Theis, 2018)
Endosymbiosis	(sometimes also referred to more precisely as endocytobiosis; Nardon & Charles, 2001) –a symbiotic relationship in which a symbiont lives inside the host cells; prominent examples are mitochondria, plastids/photosymbionts, or nitrogen fixing bacteria in plant root nodules. See also ectosymbiosis

Gnotobiosis	the condition in which all organisms present in a culture can be controlled, i.e., germ-free (axenic) organisms or organisms with a controlled community of symbionts. Gnotobiotic individuals may be obtained e.g., by surgical removal from the mother (vertebrates) or by surface sterilization of seeds (plants) and subsequent handling in a sterile environment and possible inoculation with selected microbes (Hale, Lindsey & Hameed, 1973 ; Williams, 2014)
Holism	the theory that parts of a whole are in intimate interconnection, such that they cannot exist independently of the whole, or cannot be understood without reference to the whole, which is thus regarded as greater than the sum of its parts
Holobiont	an ecological unit of different species living together in symbiosis. The term is frequently used for the unit of a host and its associated microbiota but can be extended to larger scales. Whether or to what extent holobionts are also a unit of evolution is still a matter of debate (Douglas & Werren, 2016)
Hologenome	the combined genomes of the host and all members of its microbiota; (Rosenberg et al., 2007a ; Zilber-Rosenberg & Rosenberg, 2008)
Horizontal transmission	acquisition of the associated microbiome from the environment (e.g., Myers & Rothman, 1995 ; Roughgarden, 2019)
Host	the largest or dominant partner in a holobiont
Infochemical	a chemical compound, usually diffusible, that carries information on the environment, such as the presence of other organisms, and can be used to mediate inter- and intraspecific communication (Dicke & Sabelis, 1988)
Microbial gardening	the act of frequently releasing growth-enhancing or inhibiting chemicals or metabolites that favor the development of a microbial community beneficial to the host (see e.g., Saha & Weinberger, 2019)
Microbiome	the combined genetic information encoded by the microbiota; may also refer to the microbiota itself or the microbiota and its environment (see Marchesi & Ravel, 2015)
Microbiota	all microorganisms present in a particular environment or associated with a particular host (see Marchesi & Ravel, 2015)
Nested ecosystems	a view of ecosystems where each individual system, like a “Russian doll”, can be decomposed into smaller systems and/or considered part of a larger system, all of which still qualify as ecosystems (e.g., McFall-Ngai et al., 2013)
Phagocytosis	a process by which a eukaryotic cell ingests other cells or solid particles, e.g., the engulfing of symbionts by sponges (Leys et al., 2018)
Phycosphere	the physical envelope surrounding a phytoplankton cell; usually rich in organic matter (see Amin, Parker & Armbrust, 2012)

Phylosymbiosis	congruence in the phylogeny of different hosts and the composition of their associated microbiota (Brooks et al., 2016)
Rasputin effect	the phenomenon that commensals and mutualists can become parasitic in certain conditions (Overstreet & Lotz, 2016); after the Russian monk Rasputin who became the confidant of the Tsar of Russia, but later helped bring down the Tsar's empire during the Russian revolution
Sponge loop	sponges efficiently recycle dissolved organic matter turning it into detritus that becomes food for other consumers (De Goeij et al., 2013)
Symbiont	an organism living in symbiosis; usually refers to the smaller/microbial partners living in mutualistic relationships (see also host), but also includes organisms in commensalistic and parasitic relationships
Symbiosis	a close and lasting or recurrent (e.g., over generations) relationship between organisms living together; usually refers to mutualistic relationships, but also includes commensalism and parasitism
Vertical transmission	acquisition of the associated microbiome by a new generation of hosts from the parents (as opposed to horizontal transmission; e.g., Roughgarden, 2019)

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Fabrice Not is currently a PeerJ Academic Editor. Monica Medina is a former PeerJ Academic Editor.

Author Contributions

- Simon M. Dittami, Willem Stock, Catherine Leblanc and Fabrice Not conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Enrique Arboleda, Jean-Christophe Auguet, Arite Bigalke, Enora Briand, Paco Cárdenas, Ulisse Cardini, Johan Decelle, Aschwin H. Engelen, Damien Eveillard, Claire M.M. Gachon, Sarah M. Griffiths, Tilmann Harder, Ehsan Kayal, Elena Kazamia, François H. Lallier, Mónica Medina, Ezequiel M. Marzinelli, Teresa Maria Morganti, Laura Núñez Pons, Soizic Prado, José Pintado, Mahasweta Saha, Marc-André Seloosse, Derek Skillings, Shinichi Sunagawa, Eve Toulza and Alexey Vorobev conceived and designed

the experiments, performed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

This is a review paper and does not have associated raw data or code.

REFERENCES

- Alam F, Kim TY, Kim SY, Alam SS, Pramanik P, Kim PJ, Lee YB. 2015. Effect of molybdenum on nodulation, plant yield and nitrogen uptake in hairy vetch (*Vicia villosa* Roth). *Soil Science and Plant Nutrition* **61**:664–675. DOI [10.1080/00380768.2015.1030690](https://doi.org/10.1080/00380768.2015.1030690).
- Amin SA, Green DH, Hart MC, Küpper FC, Sunda WG, Carrano CJ. 2009. Photolysis of iron-siderophore chelates promotes bacterial-algal mutualism. *Proceedings of the National Academy of Sciences of the United States of America* **106**:17071–17076.
- Amin SA, Parker MS, Armbrust EV. 2012. Interactions between diatoms and bacteria. *Microbiology and Molecular Biology Reviews* **76**:667–684.
- Anantharaman K, Brown CT, Hug LA, Sharon I, Castelle CJ, Probst AJ, Thomas BC, Singh A, Wilkins MJ, Karaoz U, Brodie EL, Williams KH, Hubbard SS, Banfield JF. 2016. Thousands of microbial genomes shed light on interconnected biogeochemical processes in an aquifer system. *Nature Communications* **7**:13219.
- Arboleda E, Hartenstein V, Martinez P, Reichert H, Sen S, Sprecher S, Bailly X. 2018. An emerging system to study photosymbiosis, brain regeneration, chronobiology, and behavior: the marine acoel *Symsagittifera roscoffensis*. *BioEssays* **40**:e1800107.
- Archibald JM. 2015. Endosymbiosis and eukaryotic cell evolution. *Current Biology* **25**:R911–R921.
- Baedke J, Fábregas-Tejeda A, Nieves Delgado A. 2020. The holobiont concept before Margulis. *Journal of Experimental Zoology Part B: Molecular and Developmental Evolution* **334**:149–155.
- Baker DM, Freeman CJ, Wong JCY, Fogel ML, Knowlton N. 2018. Climate change promotes parasitism in a coral symbiosis. *ISME Journal* **12**:921–930.
- Bang C, Dagan T, Deines P, Dubilier N, Duschl WJ, Fraune S, Hentschel U, Hirt H, Hülter N, Lachnit T, Picazo D, Pita L, Pogoreutz C, Radecker N, Saad MM, Schmitz RA, Schulenburg H, Voolstra CR, Weiland-Bräuer N. 2018. Metaorganisms in extreme environments: do microbes play a role in organismal adaptation? *Zoology* **127**:1–19.
- Baumgarten S, Simakov O, Esherick LY, Liew YJ, Lehnert EM, Michell CT, Li Y, Hambleton EA, Guse A, Oates ME, Gough J, Weis VM, Aranda M, Pringle JR, Voolstra CR. 2015. The genome of *Aiptasia*, a sea anemone model for coral symbiosis. *Proceedings of the National Academy of Sciences of the United States of America* **112**:11893–11898.

- Bell JJ, Rovellini A, Davy SK, Taylor MW, Fulton EA, Dunn MR, Bennett HM, Kandler NM, Luter HM, Webster NS. 2018. Climate change alterations to ecosystem dominance: how might sponge-dominated reefs function? *Ecology* 99:1920–1931.
- Bennett AF, Haslem A, Cheal DC, Clarke MF, Jones RN, Koehn JD, Lake PS, Lumsden LF, Lunt ID, Mackey BG, Nally RM, Menkhorst PW, New TR, Newell GR, Hara T, Quinn GP, Radford JQ, Robinson D, Watson JEM. 2009. Ecological processes: a key element in strategies for nature conservation. *Ecological Management & Restoration* 10:192–199.
- Berkelmans R, van Oppen MJH. 2006. The role of zooxanthellae in the thermal tolerance of corals: a nugget of hope for coral reefs in an era of climate change. *Proceedings of the Royal Society B: Biological Sciences* 273:2305–2312.
- Berruti A, Lumini E, Balestrini R, Bianciotto V. 2015. Arbuscular mycorrhizal fungi as natural biofertilizers: let's benefit from past successes. *Frontiers in Microbiology* 6:1559.
- Berry D, Loy A. 2018. Stable-isotope probing of human and animal microbiome function. *Trends in Microbiology* 26:999–1007.
- Björk JR, Díez-Vives C, Astudillo-García C, Archie EA, Montoya JM. 2019. Vertical transmission of sponge microbiota is inconsistent and unfaithful. *Nature Ecology & Evolution* 3:1172–1183 DOI 10.1038/s41559-019-0935-x.
- Bonaglia S, Brüchert V, Callac N, Vicenzi A, Chi Fru E, Nascimento FJA. 2017. Methane fluxes from coastal sediments are enhanced by macrofauna. *Scientific Reports* 7:13145 DOI 10.1038/s41598-017-13263-w.
- Bordron P, Latorre M, Cortés M-P, González M, Thiele S, Siegel A, Maass A, Eveillard D. 2016. Putative bacterial interactions from metagenomic knowledge with an integrative systems ecology approach. *Microbiologyopen* 5:106–117 DOI 10.1002/mbo3.315.
- Bouchard Marmen M, Kenchington E, Ardyna M, Archambault . 2017. Influence of seabird colonies and other environmental variables on benthic community structure, Lancaster Sound region, Canadian Arctic. *Journal of Marine Systems* 167:105–117 DOI 10.1016/j.jmarsys.2016.11.021.
- Bowers RM, Doud DFR, Woyke T. 2017. Analysis of single-cell genome sequences of bacteria and archaea. *Emerging Topics in Life Sciences* 1:249–255 DOI 10.1042/ETLS20160028.
- Bright M, Bulgheresi S. 2010. A complex journey: transmission of microbial symbionts. *Nature Reviews. Microbiology* 8:218–230 DOI 10.1038/nrmicro2262.
- Brooks AW, Kohl KD, Brucker RM, van Opstal EJ, Bordenstein SR. 2016. Phyllosymbiosis: relationships and functional effects of microbial communities across host evolutionary history. *PLOS Biology* 14:e2000225 DOI 10.1371/journal.pbio.2000225.
- Bulgarelli D, Schlaeppi K, Spaepen S, Ver Loren van Themaat E, Schulze-Lefert . 2013. Structure and functions of the bacterial microbiota of plants. *Annual Review of Plant Biology* 64:807–838 DOI 10.1146/annurev-arplant-050312-120106.

- Burgess SC, Baskett ML, Grosberg RK, Morgan SG, Strathmann RR. 2016. When is dispersal for dispersal? Unifying marine and terrestrial perspectives. *Biological Reviews* 91:867–882 DOI 10.1111/brv.12198.
- Burgunter-Delamare B, KleinJan H, Frioux C, Fremy E, Wagner M, Corre E, Le Salver A, Leroux C, Leblanc C, Boyen C, Siegel A, Dittami SM. 2020. Metabolic complementarity between a brown alga and associated cultivable bacteria provide indications of beneficial interactions. *Frontiers in Marine Science* 7:85.
- Burke C, Steinberg P, Rusch D, Kjelleberg S, Thomas T. 2011a. Bacterial community assembly based on functional genes rather than species. *Proceedings of the National Academy of Sciences of the United States of America* 108:14288–14293 DOI 10.1073/pnas.1101591108.
- Burke C, Thomas T, Lewis M, Steinberg P, Kjelleberg S. 2011b. Composition, uniqueness and variability of the epiphytic bacterial community of the green alga *Ulva australis*. *ISME Journal* 5:590–600 DOI 10.1038/ismej.2010.164.
- Cardini U, Bednarz VN, Naumann MS, van Hoytema N, Rix L, Foster RA, Al-Rshaidat MMD, Wild C. 2015. Functional significance of dinitrogen fixation in sustaining coral productivity under oligotrophic conditions. *Proceedings of the Royal Society B: Biological Sciences* 282:20152257 DOI 10.1098/rspb.2015.2257.
- Chiarello M, Auguet J-C, Bettarel Y, Bouvier C, Claverie T, Graham NAJ, Rieuvilleneuve F, Sucré E, Bouvier T, Villéger S. 2018. Skin microbiome of coral reef fish is highly variable and driven by host phylogeny and diet. *Microbiome* 6:147 DOI 10.1186/s40168-018-0530-4.
- Cho D-H, Ramanan R, Heo J, Lee J, Kim B-H, Oh H-M, Kim H-S. 2015. Enhancing microalgal biomass productivity by engineering a microalgal–bacterial community. *Bioresource Technology* 175:578–585 DOI 10.1016/j.biortech.2014.10.159.
- Cirri E, Pohnert G. 2019. Algae –bacteria interactions that balance the planktonic microbiome. *New Phytologist* 223:100–106 DOI 10.1111/nph.15765.
- Clay K, Holah J. 1999. Fungal endophyte symbiosis and plant diversity in successional fields. *Science (80-.)* 285:1742–1744 DOI 10.1126/science.285.5434.1742.
- Coles VJ, Stukel MR, Brooks MT, Burd A, Crump BC, Moran MA, Paul JH, Satinsky BM, Yager PL, Zielinski BL, Hood RR. 2017. Ocean biogeochemistry modeled with emergent trait-based genomics. *Science (80-.)* 358:1149–1154 DOI 10.1126/science.aan5712.
- Colin S, Coelho LP, Sunagawa S, Bowler C, Karsenti E, Bork P, Pepperkok R, de Vargas C. 2017. Quantitative 3D-imaging for cell biology and ecology of environmental microbial eukaryotes. *Elife* 6:e26066 DOI 10.7554/eLife.26066.
- Cooper MB, Kazamia E, Helliwell KE, Kudahl UJ, Sayer A, Wheeler GL, Smith AG. 2019. Cross-exchange of B-vitamins underpins a mutualistic interaction between *Ostreococcus tauri* and *Dinoroseobacter shibae*. *ISME Journal* 13:334–345 DOI 10.1038/s41396-018-0274-y.
- Costanza R, De Groot R, Sutton P, Van der Ploeg S, Anderson SJ, Kubiszewski I, Farber S, Turner RK. 2014. Changes in the global value of ecosystem services. *Global Environmental Change* 26:152–158 DOI 10.1016/j.gloenvcha.2014.04.002.

- De Goeij JM, Van Oevelen D, Vermeij MJA, Osinga R, Middelburg JJ, De Goeij AFPM, Admiraal W. 2013. Surviving in a marine desert: the sponge loop retains resources within coral reefs. *Science* (80-.) **342**:108–110 DOI [10.1126/science.1241981](https://doi.org/10.1126/science.1241981).
- De Lorgeril J, Lucasson A, Petton B, Toulza E, Montagnani C, Clerissi C, Vidal-Dupiol J, Chaparro C, Galinier R, Escoubas J-M, Haffner P, Dégremont L, Charrière GM, Lafont M, Delort A, Vergnes A, Chiarello M, Faury N, Rubio T. 2018. Immune-suppression by OsHV-1 viral infection causes fatal bacteraemia in Pacific oysters. *Nature Communications* **9**:4215 DOI [10.1038/s41467-018-06659-3](https://doi.org/10.1038/s41467-018-06659-3).
- De Schryver P, Vadstein O. 2014. Ecological theory as a foundation to control pathogenic invasion in aquaculture. *ISME Journal* **8**:2360–2368 DOI [10.1038/ismej.2014.84](https://doi.org/10.1038/ismej.2014.84).
- De Vargas C, Audic S, Henry N, Decelle J, Mahé F, Logares R, Lara E, Berney C, Bescot NLe, Probert I, Carmichael M, Poulain J, Romac S, Colin S, Aury J-M, Bittner L, Chaffron S, Dunthorn M, Engelen S. 2015. Eukaryotic plankton diversity in the sunlit ocean. *Science* (80-.) **348**:1261605 DOI [10.1126/science.1261605](https://doi.org/10.1126/science.1261605).
- De Vienne DM, Refrégier G, López-Villavicencio M, Tellier A, Hood ME, Giraud T. 2013. Cospeciation vs host-shift speciation: methods for testing, evidence from natural associations and relation to coevolution. *New Phytologist* **198**:347–385 DOI [10.1111/nph.12150](https://doi.org/10.1111/nph.12150).
- Decelle J, Colin S, Foster RA. 2015. Marine protists. In: Ohtsuka S, Suzuki T, Horiguchi T, Suzuki N, Not F, eds. *Marine Protists*. Tokyo: Springer Japan, 465–500.
- Decelle J, Stryhanyuk H, Gallet B, Veronesi G, Schmidt M, Balzano S, Marro S, Uwizeye C, Jouneau P-H, Lupette J, Jouhet J, Maréchal E, Schwab Y, Schieber NL, Tucoulou R, Richnow H, Finazzi G, Musat N. 2019. Algal remodeling in a ubiquitous planktonic photosymbiosis. *Current Biology* **29**:968–978 DOI [10.1016/j.cub.2019.01.073](https://doi.org/10.1016/j.cub.2019.01.073).
- Dicke M, Sabelis MW. 1988. Infochemical terminology: based on cost-benefit analysis rather than origin of compounds? *Functional Ecology* **2**:131–139 DOI [10.2307/2389687](https://doi.org/10.2307/2389687).
- Dickie IA, Bufford JL, Cobb RC, Desprez-Loustau M-L, Grelet G, Hulme PE, Klironomos J, Makiola A, Nuñez MA, Pringle A, Thrall PH, Tourtellot SG, Waller L, Williams NM. 2017. The emerging science of linked plant-fungal invasions. *New Phytologist* **215**:1314–1332 DOI [10.1111/nph.14657](https://doi.org/10.1111/nph.14657).
- Dini-Andreote F, Stegen JC, van Elsas JD, Salles JF. 2015. Disentangling mechanisms that mediate the balance between stochastic and deterministic processes in microbial succession. *Proceedings of the National Academy of Sciences of the United States of America* **112**:E1326–E1332 DOI [10.1073/pnas.1414261112](https://doi.org/10.1073/pnas.1414261112).
- Dittami SM, Duboscq-Bidot LL, Perennou M, Gobet AA, Corre E, Boyen C, Tonon T. 2016. Host–microbe interactions as a driver of acclimation to salinity gradients in brown algal cultures. *ISME Journal* **10**:51–63 DOI [10.1038/ismej.2015.104](https://doi.org/10.1038/ismej.2015.104).
- Dittami SM, Eveillard D, Tonon T. 2014. A metabolic approach to study algal–bacterial interactions in changing environments. *Molecular Ecology* **23**:1656–1660 DOI [10.1111/mec.12670](https://doi.org/10.1111/mec.12670).

- Douglas AE, Werren JH. 2016. Holes in the hologenome: why host-microbe symbioses are not holobionts. *MBio* 7:e02099.
- Dubilier N, Bergin C, Lott C. 2008. Symbiotic diversity in marine animals: the art of harnessing chemosynthesis. *Nature Reviews. Microbiology* 6:725–740 DOI 10.1038/nrmicro1992.
- Duperron S, Halary S, Lorion J, Sibuet M, Gaill F. 2008. Unexpected co-occurrence of six bacterial symbionts in the gills of the cold seep mussel *Idas* sp. (Bivalvia: Mytilidae). *Environmental Microbiology* 10:433–445 DOI 10.1111/j.1462-2920.2007.01465.x.
- Egan S, Gardiner M. 2016. Microbial dysbiosis: rethinking disease in marine ecosystems. *Frontiers in Microbiology* 7:991.
- Epilov DZ, Batterman SA, Hedin LO, Leake JR, Smith LM, Beerling DJ. 2017. N₂-fixing tropical legume evolution: a contributor to enhanced weathering through the Cenozoic? *Proceedings of the Royal Society B: Biological Sciences* 284:20170370 DOI 10.1098/rspb.2017.0370.
- Faith JJ, Ahern PP, Ridaura VK, Cheng J, Gordon JL. 2014. Identifying gut microbe-host phenotype relationships using combinatorial communities in gnotobiotic mice. *Science Translational Medicine* 6:220ra11 DOI 10.1126/scitranslmed.3008051.
- Falkowski PG, Fenchel T, Delong EF. 2008. The microbial engines that drive Earth's biogeochemical cycles. *Science* 320:1034–1039 DOI 10.1126/science.1153213.
- Faure D, Simon J-C, Heulin T. 2018. Holobiont: a conceptual framework to explore the eco-evolutionary and functional implications of host-microbiota interactions in all ecosystems. *New Phytologist* 218:1321–1324 DOI 10.1111/nph.15199.
- Fiore CL, Jarett JK, Olson ND, Lesser MP. 2010. Nitrogen fixation and nitrogen transformations in marine symbioses. *Trends in Microbiology* 18:455–463 DOI 10.1016/j.tim.2010.07.001.
- Franzenburg S, Walter J, Kunzel S, Wang J, Baines JF, Bosch TCG, Fraune S. 2013. Distinct antimicrobial peptide expression determines host species-specific bacterial associations. *Proceedings of the National Academy of Sciences of the United States of America* 110:E3730–E3738 DOI 10.1073/pnas.1304960110.
- Funkhouser LJ, Bordenstein SR. 2013. Mom knows best: the universality of maternal microbial transmission. *PLOS Biology* 11:e1001631 DOI 10.1371/journal.pbio.1001631.
- Geier B, Sogin EM, Michellod D, Janda M, Kompauer M, Spengler B, Dubilier N, Liebeke M. 2020. Spatial metabolomics of in situ host-microbe interactions at the micrometre scale. *Nature Microbiology* 5:498–510.
- Gilbert SF, McDonald E, Boyle N, Buttino N, Gyi L, Mai M, Prakash N, Robinson J. 2010. Symbiosis as a source of selectable epigenetic variation: taking the heat for the big guy. *Philosophical Transactions of the Royal Society B: Biological Sciences* 365:671–678 DOI 10.1098/rstb.2009.0245.
- Godoy O, Bartomeus I, Rohr RP, Saavedra S. 2018. Towards the integration of niche and network theories. *Trends in Ecology & Evolution* 33:287–300 DOI 10.1016/j.tree.2018.01.007.
- Gregory AC, Zayed AA, Conceição Neto N, Temperton B, Bolduc B, Alberti A, Ardyna M, Arkhipova K, Carmichael M, Cruaud C, Dimier C, Domínguez-Huerta G,

- Ferland J, Kandels S, Liu Y, Marec C, Pesant S, Picheral M, Pisarev S. 2019. Marine DNA viral macro- and microdiversity from pole to pole. *Cell* 177:1109–1123 DOI 10.1016/j.cell.2019.03.040.
- Gribben PE, Nielsen S, Seymour JR, Bradley DJ, West MN, Thomas T. 2017. Microbial communities in marine sediments modify success of an invasive macrophyte. *Scientific Reports* 7:9845 DOI 10.1038/s41598-017-10231-2.
- Guimarães PR, Pires MM, Jordano P, Bascompte J, Thompson JN. 2017. Indirect effects drive coevolution in mutualistic networks. *Nature* 550:511–514 DOI 10.1038/nature24273.
- Gutleben J, Chaib De Mares M, van Elsas JD, Smidt H, Overmann J, Sipkema D. 2018. The multi-omics promise in context: from sequence to microbial isolate. *Critical Reviews in Microbiology* 44:212–229 DOI 10.1080/1040841X.2017.1332003.
- Hale MG, Lindsey DL, Hameed KM. 1973. Gnotobiotic culture of plants and related research. *Botanical Review* 39:261–273 DOI 10.1007/BF02860119.
- Hammer TJ, Sanders JG, Fierer N. 2019. Not all animals need a microbiome. *FEMS Microbiology Letters* 366:fnz117.
- Harder T, Campbell AH, Egan S, Steinberg PD. 2012. Chemical mediation of ternary interactions between marine holobionts and their environment as exemplified by the red alga *Delisea pulchra*. *Journal of Chemical Ecology* 38:442–450 DOI 10.1007/s10886-012-0119-5.
- Harris R. 2010. The L4 time-series: the first 20 years. *Journal of Plankton Research* 32:577–583 DOI 10.1093/plankt/fbq021.
- Hartmann AC, Petras D, Quinn RA, Protsyuk I, Archer FI, Ransome E, Williams GJ, Bailey BA, Vermeij MJA, Alexandrov T, Dorrestein PC, Rohwer FL. 2017. Meta-mass shift chemical profiling of metabolomes from coral reefs. *Proceedings of the National Academy of Sciences of the United States of America* 114:11685–11690 DOI 10.1073/pnas.1710248114.
- Hoitink H, Boehm M. 1999. Biocontrol within the context of soil microbial communities: a substrate-dependent phenomenon. *Annual Review of Phytopathology* 37:427–446 DOI 10.1146/annurev.phyto.37.1.427.
- Hollants J, Leliaert F, Verbruggen H, De Clerck O, Willems A. 2013. Host specificity and coevolution of *Flavobacteriaceae* endosymbionts within the siphonous green seaweed *Bryopsis*. *Molecular Phylogenetics and Evolution* 67:608–614 DOI 10.1016/j.ympev.2013.02.025.
- Holt EA, Miller SW. 2010. Bioindicators: using organisms to measure environmental impacts. *Nature Education Knowledge* 3:8–13.
- Kavimandan SK. 1985. *Root nodule bacteria to improve yield of wheat (Triticum aestivum L.)*. Heidelberg: Springer.
- Kazamia E, Aldridge DC, Smith AG. 2012. Synthetic ecology –A way forward for sustainable algal biofuel production? *Journal of Biotechnology* 162:163–169 DOI 10.1016/j.jbiotec.2012.03.022.

- Kazamia E, Helliwell KE, Purton S, Smith AG. 2016. How mutualisms arise in phytoplankton communities: building eco-evolutionary principles for aquatic microbes. *Ecology Letters* 19:810–822 DOI 10.1111/ele.12615.
- Kelty MO, Cook CB. 1976. Survival during starvation of symbiotic, aposymbiotic, and non-symbiotic *Hydra*. In: *Coelenterate ecology and behavior*. Boston: Springer, 409–414.
- Kessler RW, Weiss A, Kuegler S, Hermes C, Wichard T. 2018. Macroalgal–bacterial interactions: Role of dimethylsulfoniopropionate in microbial gardening by *Ulva* (Chlorophyta). *Molecular Ecology* 27:1808–1819 DOI 10.1111/mec.14472.
- Kinlan BP, Gaines SD. 2003. Propagule dispersal in marine and terrestrial environments: a community perspective. *Ecology* 84:2007–2020 DOI 10.1890/01-0622.
- Kleiner M, Dong X, Hinzke T, Wippler J, Thorson E, Mayer B, Strous M. 2018. Metaproteomics method to determine carbon sources and assimilation pathways of species in microbial communities. *Proceedings of the National Academy of Sciences of the United States of America* 115:E5576–E5584 DOI 10.1073/pnas.1722325115.
- Klinger D, Naylor R. 2012. Searching for solutions in aquaculture: charting a sustainable course. *Annual Review of Environment and Resources* 37:247–276 DOI 10.1146/annurev-environ-021111-161531.
- Kopf A, Bicak M, Kottmann R, Schnetzer J, Kostadinov I, Lehmann K, Fernandez-Guerra A, Jeanthon C, Rahav E, Ullrich M, Wichels A, Gerds G, Polymenakou P, Kotoulas G, Siam R, Abdallah RZ, Sonnenschein EC, Cariou T, O’Gara F. 2015. The ocean sampling day consortium. *Gigascience* 4:27 DOI 10.1186/s13742-015-0066-5.
- Kubo I, Hosoda K, Suzuki S, Yamamoto K, Kihara K, Mori K, Yomo T. 2013. Construction of bacteria–eukaryote synthetic mutualism. *Biosystems* 113:66–71 DOI 10.1016/j.biosystems.2013.05.006.
- Lagier JC, Armougom F, Million M, Hugon P, Pagnier I, Robert C, Bittar F, Fournous G, Gimenez G, Maraninchi M, Trape JF, Koonin EV, La Scola B, Raoult D. 2012. Microbial culturomics: Paradigm shift in the human gut microbiome study. *Clinical Microbiology and Infection* 18:1185–1193 DOI 10.1111/1469-0691.12023.
- Lanterbecq D, Rouse GW, Eeckhaut I. 2010. Evidence for cospeciation events in the host–symbiont system involving crinoids (Echinodermata) and their obligate associates, the myzostomids (Myzostomida, Annelida). *Molecular Phylogenetics and Evolution* 54:357–371 DOI 10.1016/j.ympev.2009.08.011.
- Le Chevanton M, Garnier M, Bougaran G, Schreiber N, Lukomska E, Bérard J-B, Fouilland E, Bernard O, Cadoret J-P. 2013. Screening and selection of growth-promoting bacteria for *Dunaliella* cultures. *Algal Research* 2:212–222 DOI 10.1016/j.algal.2013.05.003.
- Lebeis SL, Paredes SH, Lundberg DS, Breakfield N, Gehring J, McDonald M, Malfatti S, Glavina del Rio T, Jones CD, Tringe SG, Dangl JL. 2015. Salicylic acid modulates colonization of the root microbiome by specific bacterial taxa. *Science* (80-.) 349:860–864 DOI 10.1126/science.aaa8764.

- Lehnert EM, Burriesi MS, Pringle JR. 2012.** Developing the anemone *Aiptasia* as a tractable model for cnidarian-dinoflagellate symbiosis: the transcriptome of aposymbiotic *A. pallida*. *BMC Genomics* **13**:271 DOI [10.1186/1471-2164-13-271](https://doi.org/10.1186/1471-2164-13-271).
- Lehnert EM, Mouchka ME, Burriesi MS, Gallo ND, Schwarz JA, Pringle JR. 2014.** Extensive differences in gene expression between symbiotic and aposymbiotic cnidarians. *G3 Genes Genomes Genetics* **4**:277–295.
- Lenneman EM, Wang P, Barney BM. 2014.** Potential application of algicidal bacteria for improved lipid recovery with specific algae. *FEMS Microbiology Letters* **354**:102–110 DOI [10.1111/1574-6968.12436](https://doi.org/10.1111/1574-6968.12436).
- Leys SP, Kahn AS, Fang JKH, Kutti T, Bannister RJ. 2018.** Phagocytosis of microbial symbionts balances the carbon and nitrogen budget for the deep-water boreal sponge *Geodia barretti*. *Limnology and Oceanography* **63**:187–202 DOI [10.1002/lno.10623](https://doi.org/10.1002/lno.10623).
- Llewellyn MS, Boutin S, Hoseinifar SH, Derome N. 2014.** Teleost microbiomes: the state of the art in their characterization, manipulation and importance in aquaculture and fisheries. *Frontiers in Microbiology* **5**:207.
- Loh J, Pierson EA, Pierson LS, Stacey G, Chatterjee A. 2002.** Quorum sensing in plant-associated bacteria. *Current Opinion in Plant Biology* **5**:285–290 DOI [10.1016/S1369-5266\(02\)00274-1](https://doi.org/10.1016/S1369-5266(02)00274-1).
- Madsen EL. 2011.** Microorganisms and their roles in fundamental biogeochemical cycles. *Current Opinion in Biotechnology* **22**:456–464 DOI [10.1016/j.copbio.2011.01.008](https://doi.org/10.1016/j.copbio.2011.01.008).
- Marchesi JR, Ravel J. 2015.** The vocabulary of microbiome research: a proposal. *Microbiome* **3**:31 DOI [10.1186/s40168-015-0094-5](https://doi.org/10.1186/s40168-015-0094-5).
- Margulis L, Fester R. 1991.** *Symbiosis as a source of evolutionary innovation: speciation and morphogenesis*. Cambridge: MIT Press, 470 pp.
- Marques A, Ollevier F, Verstraete W, Sorgeloos P, Bossier P. 2006.** Gnotobiotically grown aquatic animals: opportunities to investigate host-microbe interactions. *Journal of Applied Microbiology* **100**:903–918 DOI [10.1111/j.1365-2672.2006.02961.x](https://doi.org/10.1111/j.1365-2672.2006.02961.x).
- Martin W, Baross J, Kelley D, Russell MJ. 2008.** Hydrothermal vents and the origin of life. *Nature Reviews. Microbiology* **6**:805–814 DOI [10.1038/nrmicro1991](https://doi.org/10.1038/nrmicro1991).
- Martin-Platero AM, Cleary B, Kauffman K, Preheim SP, McGillicuddy DJ, Alm EJ, Polz MF. 2018.** High resolution time series reveals cohesive but short-lived communities in coastal plankton. *Nature Communications* **9**:266 DOI [10.1038/s41467-017-02571-4](https://doi.org/10.1038/s41467-017-02571-4).
- Martínez-Pérez C, Mohr W, Löscher CR, Dekazemacker J, Littmann S, Yilmaz P, Lehnen N, Fuchs BM, Lavik G, Schmitz RA, LaRoche J, Kuypers MMM. 2016.** The small unicellular diazotrophic symbiont, UCYN-A, is a key player in the marine nitrogen cycle. *Nature Microbiology* **1**:16163 DOI [10.1038/nmicrobiol.2016.163](https://doi.org/10.1038/nmicrobiol.2016.163).
- Marzinelli EM, Campbell AH, Valdes EZozaya, Vergés A, Nielsen S, Wernberg T, Bettignies Tde, Bennett S, Caporaso JG, Thomas T, Steinberg PD. 2015.** Continental-scale variation in seaweed host-associated bacterial communities is a function of host condition, not geography. *Environmental Microbiology* **17**:4078–4088 DOI [10.1111/1462-2920.12972](https://doi.org/10.1111/1462-2920.12972).

- Marzinelli EM, Qiu Z, Dafforn KA, Johnston EL, Steinberg PD, Mayer-Pinto M. 2018. Coastal urbanisation affects microbial communities on a dominant marine holobiont. *Biofilms and Microbiomes* 4:1 DOI 10.1038/s41522-017-0044-z.
- McFall-Ngai MJ. 2014. The importance of microbes in animal development: lessons from the squid-*Vibrio* symbiosis. *Annual Review of Microbiology* 68:177–194 DOI 10.1146/annurev-micro-091313-103654.
- McFall-Ngai M, Hadfield MG, Bosch TCG, Carey HV, Domazet-Lošo T, Douglas AE, Dubilier N, Eberl G, Fukami T, Gilbert SF, Hentschel U, King N, Kjelleberg S, Knoll AH, Kremer N, Mazmanian SK, Metcalf JL, Nealson K, Pierce NE. 2013. Animals in a bacterial world, a new imperative for the life sciences. *Proceedings of the National Academy of Sciences of the United States of America* 110:3229–3236 DOI 10.1073/pnas.1218525110.
- Meng A, Marchet C, Corre E, Peterlongo P, Alberti A, Da Silva C, Wincker P, Pelletier E, Probert I, Decelle J, Le Crom S, Not F, Bittner L. 2018. A de novo approach to disentangle partner identity and function in holobiont systems. *Microbiome* 6:105 DOI 10.1186/s40168-018-0481-9.
- Mereschkowski C. 1905. Über Natur und Ursprung der Chromatophoren im Pflanzenreiche. *Biologisches Centralblatt* 25:593–604.
- Meyer-Abich A. 1943. Beiträge zur Theorie der Evolution der Organismen. I. Das typologische Grundgesetz und seine Folgerungen für Phylogenie und Entwicklungsphysiologie [Contributions to the evolutionary theory of organisms: I. The basic typological law and its implications]. *Acta Biotheoretica* 7:1–80.
- Middelboe M, Brussaard CPD. 2017. Marine viruses: key players in marine ecosystems. *Viruses* 9:302 DOI 10.3390/v9100302.
- Millennium Ecosystem Assessment Panel T. 2005. *Ecosystems and human well-being : synthesis / millennium ecosystem assessment*. Washington: Island press, 1–155.
- Mitra A, Flynn KJ, Burkholder JM, Berge T, Calbet A, Raven JA, Granéli E, Glibert PM, Hansen PJ, Stoecker DK, Thingstad F, Tillmann U, Våge S, Wilken S, Zubkov MV. 2013. The role of mixotrophic protists in the biological carbon pump. *Biogeosciences Discuss* 10:13535–13562.
- Momose T, Concordet J-P. 2016. Diving into marine genomics with CRISPR/Cas9 systems. *Marine Genomics* 30:55–65 DOI 10.1016/j.margen.2016.10.003.
- Moran NA, Sloan DB. 2015. The hologenome concept: helpful or hollow? *PLOS Biology* 13:e1002311 DOI 10.1371/journal.pbio.1002311.
- Musat N, Musat F, Weber PK, Pett-Ridge J. 2016. Tracking microbial interactions with NanoSIMS. *Current Opinion in Biotechnology* 41:114–121 DOI 10.1016/j.copbio.2016.06.007.
- Myers JH, Rothman LE. 1995. Virulence and transmission of infectious diseases in humans and insects: evolutionary and demographic patterns. *TREE* 10:194–198.
- Nardon P, Charles H. 2001. Morphological aspects of symbiosis. In: *In Symbiosis*. Dordrecht: Kluwer Academic Publishers, 13–44.
- Nemergut DR, Schmidt SK, Fukami T, O'Neill SP, Bilinski TM, Stanish LF, Knelman JE, Darcy JL, Lynch RC, Wickey P, Ferrenberg S. 2013. Patterns and processes

- of microbial community assembly. *Microbiology and Molecular Biology Reviews* 77:342–356 DOI [10.1128/MMBR.00051-12](https://doi.org/10.1128/MMBR.00051-12).
- Neufeld KM, Kang N, Bienenstock J, Foster JA. 2011. Reduced anxiety-like behavior and central neurochemical change in germ-free mice. *Neurogastroenterol. Motil* 23:255–264 DOI [10.1111/j.1365-2982.2010.01620.x](https://doi.org/10.1111/j.1365-2982.2010.01620.x).
- Nichols D, Cahoon N, Trakhtenberg EM, Pham L, Mehta A, Belanger A, Kanigan T, Lewis K, Epstein SS. 2010. Use of ichip for high-throughput in situ cultivation of uncultivable microbial species. *Applied and Environmental Microbiology* 76:2445–2450 DOI [10.1128/AEM.01754-09](https://doi.org/10.1128/AEM.01754-09).
- Not F, Probert I, Gerikas Ribeiro C, Crenn K, Guillou L, Jeanthon C, Vaultot D. 2016. Photosymbiosis in marine pelagic environments. In: *The marine microbiome*. Cham: Springer International Publishing, 305–332.
- Ochsenkühn MA, Schmitt-Kopplin P, Harir M, Amin SA. 2018. Coral metabolite gradients affect microbial community structures and act as a disease cue. *Communications Biology* 1:184 DOI [10.1038/s42003-018-0189-1](https://doi.org/10.1038/s42003-018-0189-1).
- Ohdera AH, Abrams MJ, Ames CL, Baker DM, Suescún-Bolívar LP, Collins AG, Freeman CJ, Gamero-Mora E, Goulet TL, Hofmann DK, Jaimes-Becerra A, Long PF, Marques AC, Miller LA, Mydlarz LD, Morandini AC, Newkirk CR, Putri SP, Samson JE. 2018. Upside-down but headed in the right direction: review of the highly versatile *Cassiopea xamachana* system. *Frontiers in Ecology and Evolution* 6:35 DOI [10.3389/fevo.2018.00035](https://doi.org/10.3389/fevo.2018.00035).
- O'Malley MA. 2017. From endosymbiosis to holobionts: evaluating a conceptual legacy. *Journal of Theoretical Biology* 434:34–41 DOI [10.1016/j.jtbi.2017.03.008](https://doi.org/10.1016/j.jtbi.2017.03.008).
- Ovaskainen O, Tikhonov G, Norberg A, Guillaume Blanchet F, Duan L, Dunson D, Roslin T, Abrego N. 2017. How to make more out of community data? A conceptual framework and its implementation as models and software. *Ecology Letters* 20:561–576 DOI [10.1111/ele.12757](https://doi.org/10.1111/ele.12757).
- Overstreet RM, Lotz JM. 2016. Host–symbiont relationships: understanding the change from guest to pest. In: Hurst CJ, ed. *The rasputin effect: when commensals and symbionts become parasitic*. Cham: Springer, 27–64.
- Pan J, Stephenson AL, Kazamia E, Huck WTS, Dennis JS, Smith AG, Abell C. 2011. Quantitative tracking of the growth of individual algal cells in microdroplet compartments. *Integrative Biology* 3:1043 DOI [10.1039/c1ib00033k](https://doi.org/10.1039/c1ib00033k).
- Peek AS, Feldman RA, Lutz RA, Vrijenhoek RC. 1998. Cospeciation of chemosymbiotic bacteria and deep sea clams. *Proceedings of the National Academy of Sciences of the United States of America* 95:9962–9966 DOI [10.1073/pnas.95.17.9962](https://doi.org/10.1073/pnas.95.17.9962).
- Peixoto RS, Rosado PM, Leite D.C. de, A, Rosado AS, Bourne DG. 2017. Beneficial microorganisms for corals (BMC): proposed mechanisms for coral health and resilience. *Frontiers in Microbiology* 8:341.
- Pesant S, Not F, Picheral M, Kandels-Lewis S, Le Bescot N, Gorsky G, Iudicone D, Karsenti E, Speich S, Troublé R, Dimier C, Searson S, Acinas SG, Bork P, Boss E, Bowler C, De Vargas C, Follows M, Gorsky G. 2015. Open science resources

- p>for the discovery and analysis of Tara Oceans data.
- Scientific Data*
- 2:150023
-
- [DOI 10.1038/sdata.2015.23](https://doi.org/10.1038/sdata.2015.23)
- .
- Petersen JM, Kemper A, Gruber-Vodicka H, Cardini U, van der Geest M, Kleiner M, Bulgheresi S, Mußmann M, Herbold C, Seah BKB, Antony CP, Liu D, Belitz A, Weber M. 2016.** Chemosynthetic symbionts of marine invertebrate animals are capable of nitrogen fixation. *Nature Microbiology* 2:16195.
- Pita L, Rix L, Slaby BM, Franke A, Hentschel U. 2018.** The sponge holobiont in a changing ocean: from microbes to ecosystems. *Microbiome* 6:46
[DOI 10.1186/s40168-018-0428-1](https://doi.org/10.1186/s40168-018-0428-1).
- Pollock FJ, McMinds R, Smith S, Bourne DG, Willis BL, Medina M, Thurber RV, Zaneveld JR. 2018.** Coral-associated bacteria demonstrate phyllosymbiosis and cophylogeny. *Nature Communications* 9:4921 [DOI 10.1038/s41467-018-07275-x](https://doi.org/10.1038/s41467-018-07275-x).
- Ponnudurai R, Kleiner M, Sayavedra L, Petersen JM, Moche M, Otto A, Becher D, Takeuchi T, Satoh N, Dubilier N, Schweder T, Markert S. 2017.** Metabolic and physiological interdependencies in the *Bathymodiolus azoricus* symbiosis. *ISME Journal* 11:463–477 [DOI 10.1038/ismej.2016.124](https://doi.org/10.1038/ismej.2016.124).
- Porras-Alfaro A, Bayman P. 2011.** Hidden fungi, emergent properties: endophytes and microbiomes. *Annual Review of Phytopathology* 49:291–315
[DOI 10.1146/annurev-phyto-080508-081831](https://doi.org/10.1146/annurev-phyto-080508-081831).
- Quinn RA, Vermeij MJA, Hartmann AC, Galtier d’Auriac I, Benler S, Haas A, Quistad SD, Lim YW, Little M, Sandin S, Smith JE, Dorrestein PC, Rohwer F. 2016.** Metabolomics of reef benthic interactions reveals a bioactive lipid involved in coral defence. *Proceedings of the Royal Society B: Biological Sciences* 283:20160469
[DOI 10.1098/rspb.2016.0469](https://doi.org/10.1098/rspb.2016.0469).
- Raina J-B, Clode PL, Cheong S, Bougoure J, Kilburn MR, Reeder A, Forêt S, Stat M, Beltran V, Thomas-Hall P, Tapiolas D, Motti CM, Gong B, Pernice M, Marjo CE, Seymour JR, Willis BL, Bourne DG. 2017.** Subcellular tracking reveals the location of dimethylsulfoniopropionate in microalgae and visualises its uptake by marine bacteria. *Elife* 6:e23008 [DOI 10.7554/eLife.23008](https://doi.org/10.7554/eLife.23008).
- Raina J-B, Tapiolas D, Willis BL, Bourne DG. 2009.** Coral-associated bacteria and their role in the biogeochemical cycling of sulfur. *Applied and Environmental Microbiology* 75:3492–3501 [DOI 10.1128/AEM.02567-08](https://doi.org/10.1128/AEM.02567-08).
- Rix L, de Goeij JM, van Oevelen D, Struck U, Al-Horani FA, Wild C, Naumann MS. 2017.** Differential recycling of coral and algal dissolved organic matter via the sponge loop. *Functional Ecology* 31:778–789 [DOI 10.1111/1365-2435.12758](https://doi.org/10.1111/1365-2435.12758).
- Rohwer F, Seguritan V, Azam F, Knowlton N. 2002.** Diversity and distribution of coral-associated bacteria. *Marine Ecology Progress Series* 243:1–10
[DOI 10.3354/meps243001](https://doi.org/10.3354/meps243001).
- Rolland J-L, Stien D, Sanchez-Ferandin S, Lami R. 2016.** Quorum sensing and quorum quenching in the phycosphere of phytoplankton: a case of chemical interactions in ecology. *Journal of Chemical Ecology* 42:1201–1211 [DOI 10.1007/s10886-016-0791-y](https://doi.org/10.1007/s10886-016-0791-y).

- Rosenberg E, Koren O, Reshef L, Efrony R, Zilber-Rosenberg I. 2007a. The role of microorganisms in coral health, disease and evolution. *Nature Reviews. Microbiology* 5:355–362 DOI 10.1038/nrmicro1635.
- Rosenberg E, Koren O, Reshef L, Efrony R, Zilber-Rosenberg I. 2007b. The hologenome theory disregards the coral holobiont: reply from Rosenberg others. *Nature Reviews. Microbiology* 5:826–826.
- Rosenberg E, Sharon G, Atad I, Zilber-Rosenberg I. 2010. The evolution of animals and plants via symbiosis with microorganisms. *Environmental Microbiology Reports* 2:500–506 DOI 10.1111/j.1758-2229.2010.00177.x.
- Rosenberg E, Zilber-Rosenberg I. 2018. The hologenome concept of evolution after 10 years. *Microbiome* 6:78 DOI 10.1186/s40168-018-0457-9.
- Roughgarden J. 2020. Holobiont evolution: mathematical model with vertical vs. horizontal microbiome transmission. *Philosophy, Theory, and Practice in Biology* 12:1–24 DOI 10.3998/ptpbio.16039257.0012.002.
- Rubin-Blum M, Antony CP, Sayavedra L, Martínez-Pérez C, Birgel D, Peckmann J, Wu Y-C, Cardenas P, MacDonald I, Marcon Y, Sahling H, Hentschel U, Dubilier N. 2019. Fueled by methane: deep-sea sponges from asphalt seeps gain their nutrition from methane-oxidizing symbionts. *ISME Journal* 13:1209–1225 DOI 10.1038/s41396-019-0346-7.
- Saha M, Berdalet E, Carotenuto Y, Fink P, Harder T, John U, Not F, Pohnert G, Potin P, Selander E, Vyverman W, Wichard T, Zupo V, Steinke M. 2019. Using chemical language to shape future marine health. *Frontiers in Ecology and the Environment* 17:530–537 DOI 10.1002/fee.2113.
- Saha M, Weinberger F. 2019. Microbial gardening by a seaweed holobiont: Surface metabolites attract protective and deter pathogenic epibacterial settlement. *Journal of Ecology* 107:2255–2265 DOI 10.1111/1365-2745.13193.
- Segev E, Wyche TP, Kim KH, Petersen J, Ellebrandt C, Vlamakis H, Barteneva N, Paulson JN, Chai L, Clardy J, Kolter R. 2016. Dynamic metabolic exchange governs a marine algal–bacterial interaction. *Elife* 5:e17473 DOI 10.7554/eLife.17473.
- Selosse M-A, Bessis A, Pozo MJ. 2014. Microbial priming of plant and animal immunity: symbionts as developmental signals. *Trends in Microbiology* 22:607–613 DOI 10.1016/j.tim.2014.07.003.
- Seyedsayamdost MR, Case RJ, Kolter R, Clardy J. 2011. The Jekyll-and-Hyde chemistry of *Phaeobacter gallaeciensis*. *Nature Chemistry* 3:331–335 DOI 10.1038/nchem.1002.
- Seymour JR, Amin SA, Raina J-B, Stocker R. 2017. Zooming in on the phycosphere: the ecological interface for phytoplankton–bacteria relationships. *Nature Microbiology* 2:17065 DOI 10.1038/nmicrobiol.2017.65.
- Shapira M. 2016. Gut microbiotas and host evolution: scaling up symbiosis. *Trends in Ecology & Evolution* 31:539–549 DOI 10.1016/j.tree.2016.03.006.
- Shreiner AB, Kao JY, Young VB. 2015. The gut microbiome in health and in disease. *Current Opinions in Gastroenterology* 31:69–75 DOI 10.1097/MOG.000000000000139.

- Singh Y, Ahmad J, Musarrat J, Ehtesham NZ, Hasnain SE. 2013. Emerging importance of holobionts in evolution and in probiotics. *Gut Pathogens* 5:12 DOI 10.1186/1757-4749-5-12.
- Skillings D. 2016. Holobionts and the ecology of organisms: multi-species communities or integrated individuals? *Biology & Philosophy* 31:875–892 DOI 10.1007/s10539-016-9544-0.
- Smriga S, Sandin SA, Azam F. 2010. Abundance, diversity, and activity of microbial assemblages associated with coral reef fish guts and feces. *FEMS Microbiology Ecology* 73:31–42.
- Spoerner M, Wichard T, Bachhuber T, Stratmann J, Oertel W. 2012. Growth and thallus morphogenesis of *Ulva mutabilis* (Chlorophyta) depends on a combination of two bacterial species excreting regulatory factors. *Journal of Phycology* 48:1433–1447 DOI 10.1111/j.1529-8817.2012.01231.x.
- Stegen JC, Lin X, Fredrickson JK, Chen X, Kennedy DW, Murray CJ, Rockhold ML, Konopka A. 2013. Quantifying community assembly processes and identifying features that impose them. *ISME Journal* 7:2069–2079 DOI 10.1038/ismej.2013.93.
- Theis KR. 2018. Hologenomics: systems-level host biology. *mSystems* 3:e00164–17 DOI 10.1128/mSystems.00164-17.
- Thompson AW, Foster RA, Krupke A, Carter BJ, Musat N, Vaultot D, Kuypers MMM, Zehr JP. 2012. Unicellular cyanobacterium symbiotic with a single-celled eukaryotic alga. *Science (80-.)* 337:1546–1550 DOI 10.1126/science.1222700.
- Thompson LR, Sanders JG, McDonald D, Amir A, Ladau J, Locey KJ, Prill RJ, Tripathi A, Gibbons SM, Ackermann G, Navas-Molina JA, Janssen S, Kopylova E, Vázquez-Baeza Y, González A, Morton JT, Mirarab S, Xu ZZech, Jiang L. 2017. A communal catalogue reveals Earth’s multiscale microbial diversity. *Nature* 551:457 DOI 10.1038/nature24621.
- Tolstoy LN. 1878. Анна Каренина. Moscow: Russian Bulletin. 1–864 pp.
- Troussellier M, Escalas A, Bouvier T, Mouillot D. 2017. Sustaining rare marine microorganisms: Macroorganisms as repositories and dispersal agents of microbial diversity. *Frontiers in Microbiology* 8:947 DOI 10.3389/fmicb.2017.00947.
- Vallet M, Strittmatter M, Murúa P, Lacoste S, Dupont J, Hubas C, Genta-Jouve G, Gachon CMM, Kim GH, Prado S. 2018. Chemically-mediated interactions between macroalgae, their fungal endophytes, and protistan pathogens. *Frontiers in Microbiology* 9:3161 DOI 10.3389/fmicb.2018.03161.
- Van der Heijden MGA, Martin FM, Selosse M-A, Sanders IR. 2015. Mycorrhizal ecology and evolution: the past, the present, and the future. *New Phytologist* 205:1406–1423 DOI 10.1111/nph.13288.
- Van Oppen MJH, Gates RD, Blackall LL, Cantin N, Chakravarti LJ, Chan WY, Cormick C, Crean A, Damjanovic K, Epstein H, Harrison PL, Jones TA, Miller M, Pears RJ, Peplow LM, Raftos DA, Schaffelke B, Stewart K, Torda G. 2017. Shifting paradigms in restoration of the world’s coral reefs. *Global Change Biology* 23:3437–3448 DOI 10.1111/gcb.13647.

- Van Oppen MJH, Oliver JK, Putnam HM, Gates RD. 2015. Building coral reef resilience through assisted evolution. *Proceedings of the National Academy of Sciences of the United States of America* 112:2307–2313 DOI 10.1073/pnas.1422301112.
- Venn AA, Loram JE, Douglas AE. 2008. Photosynthetic symbioses in animals. *Journal of Experimental Botany* 59:1069–1080 DOI 10.1093/jxb/erm328.
- Wallin IE. 1925. On the nature of mitochondria. IX. Demonstration of the bacterial nature of mitochondria. *The American Journal of Anatomy* 36:131–149 DOI 10.1002/aja.1000360106.
- Webster NS, Taylor MW, Behnam F, Lückner S, Rattei T, Whalan S, Horn M, Wagner M. 2010. Deep sequencing reveals exceptional diversity and modes of transmission for bacterial sponge symbionts. *Environmental Microbiology* 12:2070–2082.
- Wichard T. 2015. Exploring bacteria-induced growth and morphogenesis in the green macroalga order Ulvales (Chlorophyta). *Frontiers in Plant Science* 6:86.
- Wichard T, Beemelmans C. 2018. Role of chemical mediators in aquatic interactions across the prokaryote–eukaryote boundary. *Journal of Chemical Ecology* 44:1008–1021 DOI 10.1007/s10886-018-1004-7.
- Williams SCP. 2014. Gnotobiotics. *Proceedings of the National Academy of Sciences of the United States of America* 111:1661 DOI 10.1073/pnas.1324049111.
- Wiltshire KH, Kraberg A, Bartsch I, Boersma M, Franke H-D, Freund J, Gebühr C, Gerds G, Stockmann K, Wichels A. 2010. Helgoland Roads, North Sea: 45 years of change. *Estuaries and Coasts* 33:295–310 DOI 10.1007/s12237-009-9228-y.
- Wolfowicz I, Baumgarten S, Voss PA, Hambleton EA, Voolstra CR, Hatta M, Guse A. 2016. Aiptasia sp. larvae as a model to reveal mechanisms of symbiont selection in cnidarians. *Scientific Reports* 6:32366 DOI 10.1038/srep32366.
- Woyke T, Teeling H, Ivanova NN, Huntemann M, Richter M, Gloeckner FO, Boffelli D, Anderson IJ, Barry KW, Shapiro HJ, Szeto E, Kyrpides NC, Musmann M, Amann R, Bergin C, Ruehlmann C, Rubin EM, Dubilier N. 2006. Symbiosis insights through metagenomic analysis of a microbial consortium. *Nature* 443:950–955.
- Zaneveld JR, McMinds R, Vega Thurber R. 2017. Stress and stability: applying the Anna Karenina principle to animal microbiomes. *Nature Microbiology* 2:17121 DOI 10.1038/nmicrobiol.2017.121.
- Zengler K, Palsson BO. 2012. A road map for the development of community systems (CoSy) biology. *Nature Reviews. Microbiology* 10:366–372 DOI 10.1038/nrmicro2763.
- Zheng D, Liwinski T, Elinav E. 2020. Interaction between microbiota and immunity in health and disease. *Cell Research* 30:492–506 DOI 10.1038/s41422-020-0332-7.
- Zilber-Rosenberg I, Rosenberg E. 2008. Role of microorganisms in the evolution of animals and plants: the hologenome theory of evolution. *FEMS Microbiology Reviews* 32:723–735 DOI 10.1111/j.1574-6976.2008.00123.x.